

## A COMPARATIVE INVESTIGATION OF THE PARTICLE FRACTIONS FROM HOYA, DISCHIDIA AND EUPHORBIA LATICES

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### SUMMARY

Particle size, morphology and lipid composition of the latex particle fractions of 18 *Hoya*, 3 *Dischidia* and 5 *Euphorbia* species were investigated. Of these latices only two showed in vitro triterpene biosynthesis.

Some aspects of particle biosynthesis are discussed.

### 1. INTRODUCTION

The particles of a number of rubber and terpenoid containing plant latices have diverse sizes ranging from less than 0.01  $\mu\text{m}$  in *Taraxacum kok-saghyz* (HENDRICKS et al. 1944) to 50  $\mu\text{m}$  and more in *Musa* (TRECUL 1867, LLOYD 1928). Early reports about particle diameters in several plant latices are only dealing with minimum and maximum sizes determined with a light microscope (JUMELLE 1903, HENRI 1908, FICKENDEY 1909, HAUSER 1927) and range in general from 0.5 to 3–4  $\mu\text{m}$ . TRUMBULL (1942) and BLONDEAU & CURTIS (1946) reported the approximate size of *Cryptostegia grandiflora* latex particles being 0.2–1.5  $\mu\text{m}$  resp. 0.6–1.4  $\mu\text{m}$  in diameter. LUCAS (1938) determined a particle diameter distribution curve of *Hevea brasiliensis* by means of ultra-violet microscopy resulting in the broad range from 0.09–2.0  $\mu\text{m}$ . With the aid of an electron microscope the existence of smaller rubber particles was demonstrated in *Hevea* (HENDRICKS et al. 1944, SCHMIDT & KELSEY 1951, VAN DEN TEMPEL 1952, SCHOON & VAN DER BIE 1955).

Indirectly particle diameter distribution curves were determined by centrifuging as was done e.g. on *Hevea* by HESSELS 1943, SCHMIDT & KELSEY 1951 and on *Funtumia elastica* by VAN DIE 1955. In addition many latices have been investigated chemically for their particle content (VAN DIE 1955, BOITEAU & RATSIMANGA 1964, PONSINET & OURISSON 1968a). Some latices appeared to be able to synthesize their particle compounds in vitro (BANDURSKY & TEAS 1957, KLEINSCHMIDT & MOTHES 1959, PONSINET & OURISSON 1967). In the present paper studies on the particle diameter frequency distribution, particle morphology and lipid composition and incorporation of acetate, mevalonate, and isopentenyl pyrophosphate into triterpenols by tapped latex of several plants are described.

## 2. MATERIALS AND METHODS

Species of *Hoya*, *Dischidia*, and *Euphorbia* were cultivated in the green house. After incision with a razor blade latex was collected with 20 or 50  $\mu$ l capillaries. Aliquots of 20  $\mu$ l latex were diluted with 0.5 ml of a phosphate buffer pH 8.1 of a suitable strength (table 1) and chromatographed on a Sephadex G-100 column (30  $\times$  0.9 cm). Parts of the eluted particle suspension were fixed with  $\text{OsO}_4$  and investigated electron optically as described before (GROENEVELD 1976).

Total terpenoid extracts of Sephadex filtrated particle suspensions were chromatographed on thin layer as described before (GROENEVELD 1976).

The dry weight of the particle fraction was determined after adding 0.3–0.5 ml 1.5 M sucrose containing 1% acetic acid to aliquots of 150–200  $\mu$ l latex and subsequent freezing; the coagulated suspension was centrifuged at 60,000 g, the top layer squeezed, washed with water, and dried to a constant weight at 60°C.

*Incorporation experiments.* 2.5  $\mu$ Ci Na-acetate 2- $\text{C}^{14}$  in 50  $\mu$ l (specific activity 55.8 m Ci/mmol) was added to 0.2 ml freshly tapped latex and incubated at 27°C during 16 hours. Light petroleum extracts were made as described before (GROENEVELD 1976) and saponified with 5% KOH in ethanol/benzene (9/1, v/v) during 1½ hours at 90°C. Unsaponifiable compounds were extracted with light petroleum (b.r. 40–60°C) and chromatographed on columns of aluminium oxide (Merck, Aluminium Oxid standardisiert, Aktivitätsstufe II–III Brockmann) by a stepwise increase of peroxide free anhydrous diethyl ether in light petroleum (E/P) (modified after GOAD & GOODWIN 1966). The fraction eluted with 10% E/P contained squalene and farnesol (partly), 30% E/P the triterpenols (4,4 dimethylsterols) and farnesol (partly), 50% E/P the sterolfraction (4-desmethylsterols).

The triterpenol and sterol fractions were acetylated in a mixture of acetic anhydride/pyridine 1/2 (90°C during 2 hours) extracted with light petroleum and chromatographed on aluminium oxide. The acetates were eluted with 10% E/P and chromatographed on thin layer plates (silicagel G impregnated with 15%  $\text{AgNO}_3$ , w/w) developed in benzene/petroleum ether, (2/3, v/v in order to separate the triterpene acetates from farneso acetate. Rf values:  $\beta$ -amyrine acetate 0.5, lupeol acetate 0.2, stigmasterol acetate 0.45, farnesol acetate 0.02, geraniol acetate 0.05. Radioactive triterpene- and sterol acetates were scraped off and counted in a liquid scintillation counter.

## 3. RESULTS

The buffer used in gel filtration of the latex of *Hoya australis* also appeared to be useful for the isolation of the particle fraction of a few other latices. Of some *Hoya* species, however, the latices flocculated during or after gel filtration, or during their fixation in  $\text{OsO}_4$ . This could be prevented by an increase of buffer strength.

As far as the *Hoya* species are concerned, the buffer concentrations given in

Table 1. Qualitative lipid composition and average diameter of latex particles in *Hoya*, *Dischidia* and *Euphorbia* latices. Buffer concentration near the minimal buffer strength necessary to prevent flocculation during gel filtration.

	Triterpenol	Triterpene acetates	Triterpene cinnamates	Triterpenes esterified with fatty acids	Average particle diameter ( $\mu\text{m}$ )	Standard deviation	Number of measured particles	Mol. buffer
<i>H. bella</i> Hook.	+	+	+	+	0.41	0.04	(554)	0.10
<i>H. coronaria</i> Blume	+	+	+	—	0.36	0.05	(279)	0.10
<i>H. shepherdii</i> Short	+	+	+	—	0.35	0.04	(444)	0.10
<i>H. australis</i> R.Br. ex Traill	+	+	+	tr.	0.34	0.05	(320)	0.12
<i>H. ovalifolia</i> Wight & Arn.	+	+	+	—	0.34	0.04	(332)	0.12
<i>H. cinnamomifolia</i> Hook.	+	+	+	—	0.33	0.05	(559)	0.12
<i>H. spec.</i>	+	—	+	—	0.32	0.06	(232)	0.12
<i>H. bandaensis</i> Schlechter	+	+	+	—	0.28	0.04	(326)	0.15
<i>H. diversifolia</i> Blume	tr.	+	+	—	0.28	0.04	(241)	0.15
<i>H. macrophylla</i> Blume	+	+	+	—	0.27	0.04	(567)	0.15
<i>H. obovata</i> Decne	—	+	+	—	0.26	0.04	(170)	0.20
<i>H. lacunosa</i> Blume	+	+	—	—	0.26	0.05	(235)	0.20
<i>H. fraterna</i> Blume	tr.	+	+	—	0.25	0.03	(276)	0.20
<i>H. latifolia</i> Don.	+	+	+	—	0.25	0.04	(191)	0.20
<i>H. crassipes</i> Turcz.	tr.	+	+	—	0.24	0.04	(227)	0.20
<i>H. longuifolia</i> Wallich	+	+	+	—	0.23	0.04	(359)	0.25
<i>H. pseudolanceolata</i> Cost.	+	tr.	+	+	0.22	0.04	(205)	0.22
<i>H. imperialis</i> Lindl.	+	+	—	+	0.20	0.03	(240)	0.25
<i>H. multiflora</i> Bl.	+	+	+	+	0.18	0.04	(235)	0.30
<i>D. membranifolia</i> (?)	+	+	—	—	0.35	0.06	(601)	0.10
<i>D. spec.</i>	+	+	—	—	0.47	0.05	(201)	0.10
<i>D. bengalensis</i> Colebr.	+	+	—	—	0.53	0.04	(345)	0.10
<i>E. balsamifera</i> Ait.	+	—	—	—	0.22	0.05	(429)	0.20
<i>E. lactea</i> Haw.	+	—	—	—	0.24	0.08	(502)	0.20
<i>E. tirucalli</i> L.	+	—	—	—	0.24	0.07	(493)	0.20
<i>E. milii</i> Desm.	+	+	—	+	0.31	0.09	(321)	0.12
<i>E. pulcherrima</i> Willd. ex Klotsch	+	+	—	+	0.28	0.06	(418)	0.2 M KCl + 0.01% Tween

table 1 may be regarded as near the minimum values needed to prevent flocculation. In these solutions particle suspensions remained stable during at least 6 hours, and also during fixation. Similar results were obtained with *Euphorbia* latices. Latex from *E. pulcherrima* flocculated immediately after dilution with phosphate buffer regardless its ionic strength and pH, but good gel filtration results could be obtained by using a 0.2 M KCl solution-containing 0.01% Tween – 80. This solution appeared to be useful in all other investigated latices too.

Latex was obtained from stem, leaves, leafstalks, young and older parts of specimen of *Hoya australis*. The triterpene particles of these samples were examined on diameter frequency distribution by scanning electron microscopy (SEM), as well as on general appearance in ultra thin sections. The particle populations of these various latex samples appeared to be very uniform throughout the plant (table 2). Like *Hoya* species, the investigated species of *Dischidia* and *Euphorbia* also have non-articulated laticifers. Here too no differences in particle morphology could be observed in latex samples of various plant parts.

All investigated *Hoya*, *Dischidia*, and *Euphorbia* species showed globular particles with a smooth surface in scanning electron microscopy. Of *Euphorbia milii* and *E. pulcherrima* some particles, especially the larger ones, were provided with a "hole" (fig. 1). This structure was observed after gel filtration in either phosphate buffer or in a 0.2 M KCl solution containing 0.01% Tween – 80. In thin sections particles with an identical structure did occur (fig. 2a, b). Here, the particle mass locally has a concave surface to which a membrane-like vesicle is attached.

In sectioned particles of *Hoya australis* sometimes a similar structure can be observed (fig. 3a, 3b), which differs from that found in the *Euphorbia* species. The concave part of the particle surface is apparently not covered with a membrane-like envelope, but this is extruded from the deformed particle, as was shown earlier (GROENEVELD 1976). Similar phenomena could be regularly observed in flocculated particle suspensions of *H. australis* (fig. 4), obtained by decreasing the buffer concentration from 0.1 M to 0.02 M phosphate. The induced flocculation had led to coalescence, accompanied by a deformation

Table 2. Average particle diameter of several latex samples of *Hoya australis*.

Site of tapping	Average diameter
All parts of the plant	0.33 $\mu\text{m}$
All parts of the plant	0.34 $\mu\text{m}$
All parts of the plant	0.33 $\mu\text{m}$
All parts of the plant	0.34 $\mu\text{m}$
Top of the stem	0.35 $\mu\text{m}$
Young leaf	0.33 $\mu\text{m}$
Young stem	0.34 $\mu\text{m}$
Old leaf	0.35 $\mu\text{m}$
Old stem	0.34 $\mu\text{m}$
Average diameter	0.34 $\mu\text{m}$

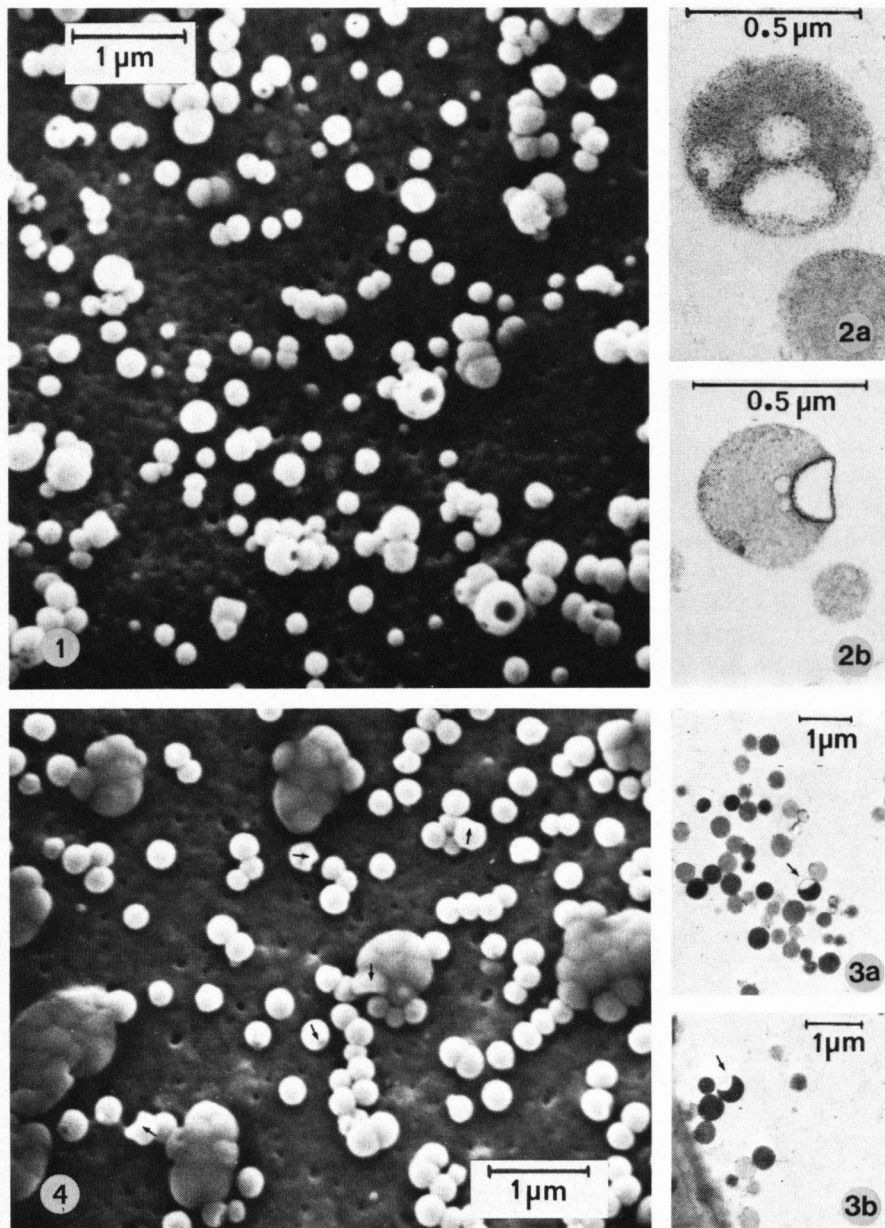


Fig. 1. Particles of *E. milii* with "holes". Phosphate buffer pH 8.0; 0.1 M,  $\text{OsO}_4$  fixation. SEM.

Fig. 2a, b. Ultra thin sections of particles of *E. milii* with membranous vesicles. Phosphate buffer pH 8.0; 0.1 M,  $\text{OsO}_4$  fixation.

Fig. 3a, b. Ultra thin sections of particles of *H. australis*. Two deformed particles between globular ones. Phosphate buffer pH 8.0; 0.1 M,  $\text{OsO}_4$  fixation.

Fig. 4. Partly coalesced particle fraction of *H. australis*, deformed particles indicated by arrows. Phosphate buffer pH 8.0; 0.1 M,  $\text{OsO}_4$  fixation. SEM.

of the globular particle shape. Such particle deformations, however, were not observed in the investigated latices of which stable suspensions were put on the millipore filter.

The results of the particle diameter frequency distribution measurements of a number of latices are given in histograms in *fig. 5*; average diameter, standard deviation, number of particles measured, and the buffer concentrations employed in gel filtration are listed in *table 1*.

Some qualitative data on the chemical composition of the various latices obtained by thin layer chromatography are also shown in *table 1*. They are in agreement with more detailed data on *Hoya* latex obtained by WARNAAR & KNOLLEMA (1976). Sterols were present as traces only, the main components being triterpene acetates in *Dischidia* and triterpene cinnamates in *Hoya* in addition to free triterpenols. Some quantitative data on several particle populations are presented in *table 3*.

Freshly tapped latex of all investigated *Hoya*'s, *Dischidia membranifolia*, and *Euphorbia balsamifera*, *E. lactea* and *E. tirucalli* were found to be unable to synthesize triterpenols from acetate *in vitro*. Even after a 16 hours' incubation period no incorporation of acetate into triterpenes or triterpene esters could be detected. Also attempts with labelled mevalonate and isopentenyl pyrophosphate were negative. Latices of *E. milii* and *E. pulcherrima*, however, showed a distinct incorporation of acetate in the triterpenol fraction as summarized in *table 4*.

#### 4. DISCUSSION

The particle populations of the investigated *Hoya* and *Dischidia* species are characterised by a narrow diameter distribution range, those of *Euphorbia*

Table 3. Dry particle content (dry weight of the particle fraction as a percentage of latex fresh weight), number of particles per  $\mu\text{l}$  latex and interfacial area (total particle surface in 1  $\mu\text{l}$  latex) of some *Euphorbia*, *Dischidia* and *Hoya* latices.

	dry particle content (%)	number of particles per $\mu\text{l}$ latex ( $\times 10^9$ )	interfacial area ( $\text{cm}^2$ )
<i>Euphorbia balsamifera</i>	7.7	11.9	19
<i>E. tirucalli</i>	18.0	12.1	23
<i>E. lactea</i>	18.0	18	36.9
<i>Dischidia spec.</i>	24.6	3.9	29.2
<i>Hoya australis</i>	7.6	3.4	11.9
<i>H. cinnamomifolia</i>	8.0	4.1	14.1
<i>H. obovata</i>	8.8	9.2	19.6
<i>H. lacunosa</i>	9.3	10	20.6
<i>H. shepherdii</i>	9.5	3.7	15.3
<i>H. coronaria</i>	12.7	4.9	20.5
<i>H. multiflora</i>	24.0	78	80.1
<i>H. imperialis</i>	28.0	68	83.4

Table 4. Incorporation of acetic acid 2-C<sup>14</sup> (sodium salt) into the triterpenol and sterol fraction of *Euphorbia milii* and *E. pulcherrima* latex. 0.2 ml latex incubated with 50 µl isotope solution (= 2.5 µCi) during 16 hours at 27°C.

	Triterpenol	Sterol
<i>E. milii</i>	72,000 dpm	1,100 dpm
<i>E. pulcherrima</i>	122,000 dpm	700 dpm

latices have a somewhat wider one and resemble those of *Funtumia elastica* (VAN DIE 1955). For *Hoya* species a uniform particle population is probably always present throughout the whole plant. The possible existence of two distinct size classes e.g. as reported for whip latex in *Cryptostegia* (0.2 and 1.1 µm, BLONDEAU & CURTIS 1946) has been carefully investigated but no evidence for it could be found.

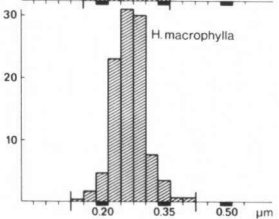
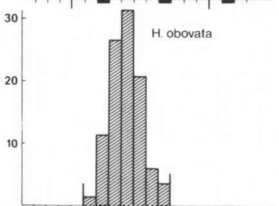
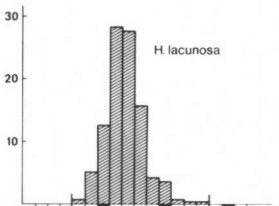
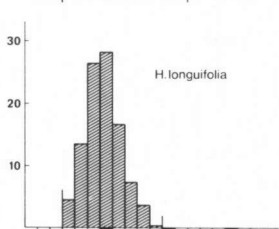
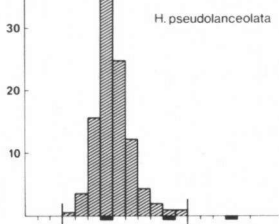
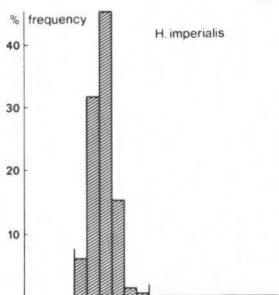
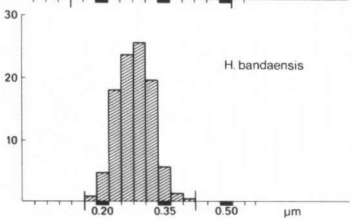
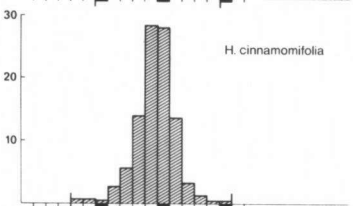
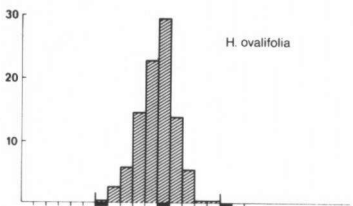
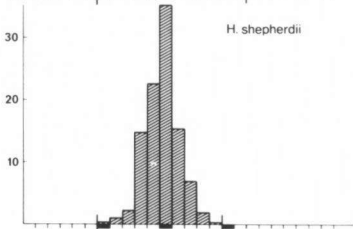
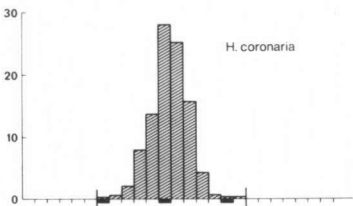
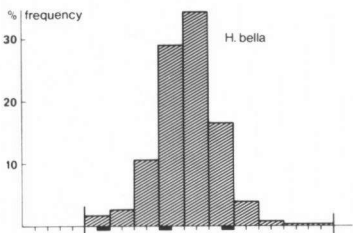
Within the genus *Hoya* an inverse correlation appeared to exist between the average particle diameter and the buffer concentration required for stabilizing the particle suspension. This supports the view that the different particle diameter distributions shown in *fig. 5* and *table 1* are characteristic features of the investigated *Hoya* latices, and not produced by different degrees of swelling caused by OsO<sub>4</sub> treatment.

In all investigated species a more or less sharply defined order of size of the smallest particles can be observed. Minimum particle size exceeds by at least a factor 4 the millipore filter grade (0.025 µm). In addition, the fluffy particle surface (inherent to the method used) gives the smallest particles a foggy image, which means that particles with a 1.5 mm diameter in a 17,000 enlargement (corresponding with 0.09 µm diameter) can be recognised as a real latex particle. Very small particles like the smallest ones (0.03 µm diameter) reported for *Hevea brasiliensis* latex (BATEMAN 1963) can not be detected with this method. The symmetric particle distributions found in the present work, however, do not suggest the existence of such very small particles in the investigated latices.

The largest particle diameter in each species could also be stated. As particle coalescence is visible in the scanning electron microscope (*fig. 4*), the suggestion of SCHOON & PHOA (1956), that the largest particles are built up from smaller ones seems improbable for *Hoya* latices.

The particle diameter frequency distributions of the latices of the investigated Asclepiadaceae are in most cases nearly symmetrical. In some species maximum frequency is found at a diameter smaller than the average one, while in other species the maximum diameter frequency exceeds the average one. All investigated species differ greatly from *Hevea brasiliensis*, which has an asymmetrical distribution curve, ranging from 0.03 µm to 1–2 µm with a maximum frequency at 0.1 µm.

No correlation could be found between particle size and particle composition in general. In most *Hoya* species triterpenyl cinnamates are present and the average particle diameter ranges from 0.18 µm to 0.41 µm. Within this range,





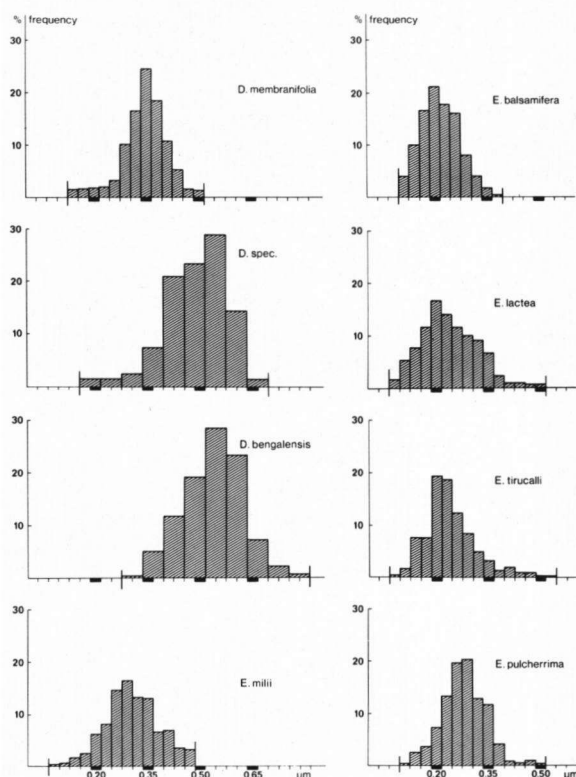


Fig. 5. Particle diameter frequency distribution in several latices; statistic interval 0.03 or 0.06  $\mu\text{m}$ . The minimum and maximum diameters are marked by vertical dashes.

however, the *Euphorbia* particles are also found, which contain either triterpenols only, or the more complex triterpene esters of *E. milii* and *E. pulcherrima*. Within the genus *Hoya*, *H. obovata* and *H. shepherdii* both only contain triterpene esters, but differ in average particle diameter. *H. bella* and *H. imperialis* both have esterified fatty acids in their particles but have nevertheless a complete different particle population.

A correlation could neither be found between  $\text{Mg}^{2+}$  or  $\text{K}^{+}$  content of the whole latex and average particle size: *H. multiflora* and *H. bella* only have small amounts of these ions but they differ greatly in particle size. A correlation between particle size and particle concentration does not exist. Latices with relatively small particles may have a high particle concentration (*H. imperialis*, *H. multiflora*) compared with those having a relatively large average particle diameter (*H. australis*). This results in the broad range of interfacial area listed in table 3.

Only in the latices of two species an in vitro triterpene biosynthesis could be demonstrated. As predominantly tetracyclic triterpenes are found in *E. milii*

latex while *E. pulcherrima* contains mainly pentacyclic triterpenes (PONSINET & OURISSON 1968a) the ability of an in vitro biosynthesis is apparently not restricted to a special type of triterpene skeleton.

Remarkably, however, these latices are characterised by a relatively high amount of potassium, and a potassium/magnesium ratio  $> 1$ , in contrast to all other investigated latices (GROENEVELD 1975). In addition both latices possess particles of which the surface has concave sites. Similar structures can be observed in the pictures of *E. pulcherrima* latex given by SCHNEPF (1964).

In *Hevea brasiliensis* the final stages of the synthesis of the main particle compound occurs at the surface of all particles (MCMULLEN & MCSWEENEY 1966). In the latex of *Euphorbia cyparissias*, however, species specific triterpenoids are synthesized in a hitherto unidentified "bottom fraction" obtained by centrifugation (PONSINET & OURISSON 1968b). Within 24 hours these newly synthesized compounds did not disperse into the supernatant in which the majority of the triterpenes were present. In tapped latex of *E. tirucalli*, *E. lactea* and in that of all mentioned *Hoya* species in vitro biosynthesis apparently does not occur. Very probably in these species the triterpene synthesizing system is not expelled with tapping. As far as the particle synthesis is concerned, the model described by POPOVICI (1926) still is an attractive one. In fixed, acid fuchsin stained sections of *Ficus carica* this author observed small rubber particles in a parietal latex vessel cytoplasm and larger ones in a central vacuole. She suggested a particle initiation in the cytoplasm and finally excretion into the vacuole, as was already suggested by TRECUL (1867) to occur in *Musa* latex. Both the narrow particle diameter frequency distribution in particle populations of *Hoya* species and the inability of in vitro triterpene biosynthesis of many latices can be explained by this model.

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#### LITERATURE

- BANDURSKI, R. S. & H. J. TEAS (1957): Rubber biosynthesis in latex of *Hevea brasiliensis*. *Plant Physiol.* **32**: 643–648.
- BATEMAN, L. (ed.) (1963): *The chemistry and physics of rubber like substances*. McLaren & Sons Ltd. London.
- BLONDEAU, R. & J. T. CURTIS (1946): Latex and Rubber Characteristics of *Cryptostegia*. *Rubber Age* **59**: 57–63.
- BOITEAU, P. & A. R. RATSIMANGA (1964): *Les triterpénoïdes en physiologie végétale et animale*. Gauthier-Villars ed. Paris.
- DIE, J. VAN (1955): A comparative study of the particle fractions from Apocynaceae latices. *Annales Bogoriensis* **2**(1): 1–124.
- FICKENDEY, E. (1909): Zur Kenntnis des Milchsafte von *Kickxia elastica*. *Tropenpflanzer* **13**: 204–209.

- GOAD L. J. & T. W. GOODWIN (1966): The biosynthesis of sterols in higher plants. *Biochem. J.* **99**: 735–746.
- GROENEVELD, H. W. (1975): Magnesium and potassium contents of latices of a number of Hoya and Euphorbia species. *Acta Bot. Neerl.* **24**: 485–488.
- (1976): Some morphological and chemical characteristics of the purified terpenoid particles of the latex of Hoya australis R.Br. ex Traill. *Acta Bot. Neerl.* **25**: 1–13.
- HAUSER, E. (1927): *Latex, sein Vorkommen, seine Gewinnung, Eigenschaften sowie technische Verwendung*. Verlag Theodor Steinkopf, Dresden und Leipzig.
- HENDRICKS, S. B., S. G. WILDMAN & H. F. MURDIE (1944): Morphology of latex particles as shown by electron micrographs. *India Rubber World* **110**: 297–300.
- HENRI, V. (1908): Recherches physico-chimiques sur le latex. In L. Morisse: “*Latex*” p. 323–336. Paris.
- HESSLS, J. H. E. (1943): *De polydispersiteit van Hevea latex*. Thesis, Delft.
- JUMELLE, H. (1903): “*Les plantes à caoutchouc et à gutta dans tous les pays chauds*”. A. Challamel, Paris, 1903.
- KLEINSCHMIDT, G. & K. MOTHES (1959): Zur Physiologie und Biosynthese der Alkaloide von Papaver somniferum. *Z. Naturforschung* **14<sup>b</sup>**: 52–56.
- LLOYD, F. E. (1928): “Colloidal aspects of Botany” in *Colloid Chemistry*, vol. II. J. Alexander ed. New York.
- LUCAS, F. F. (1938): Ultra-violet microscopy of Hevea rubber latex. *Ind. Eng. Chem.* **30**: 146–153.
- McMULLEN A. J. & G. P. MCSWEENEY (1966): Biosynthesis of Rubber. *Biochem. J.* **101**: 42–47.
- PONSINET, G. & G. OURISSON (1967): Biosynthèse in vitro des triterpènes dans le latex d'Euphorbia. *Phytochemistry* **6**: 1235–1243.
- , — (1968a): Études chimiotaxonomiques dans la famille des Euphorbiaceae III. Répartition des triterpènes dans la latex d'Euphorbia. *Phytochemistry* **7**: 89–98.
- , & — (1968b): Aspects particuliers de la biosynthèse des triterpènes dans le latex d'Euphorbia. *Phytochemistry* **7**: 757–764.
- POPOVICI, H. (1926): Contribution à l'étude cytologique des laticifères. *C.R. Acad. Sciences* **183**: 143–145.
- SCHMIDT, E. & R. H. KELSEY (1951): Creaming latex with ammonium alginate. Influence of particle size. *Industrial and Engineering Chemistry* **43**(2): 406–412.
- SCHNEPF, E. (1964): Zur Cytologie und Physiologie pflanzlicher Drüsen. 5. Elektronenmikroskopische Untersuchungen an Cyathialnektarien von Euphorbia pulcherrima in verschiedenen Funktionszuständen. *Protoplasma* **58**: 193–219.
- SCHOON, TH. G. F. & G. J. VAN DER BIE (1955): Particle size distribution in brominated Hevea latices. *J. Polymer Science* **16**: 63–88.
- , & K. L. PHOA (1956). Morphology of the Rubber particles in Natural Latices. *Archives Rubber Cultivation* **33**: 195–215.
- TEMPEL, M. VAN DEN (1952): Elektron microscopy of rubber globules in Hevea latex. *Transactions, IRI.* **28**: 303–309.
- TRÉCUL, M. A. (1867): Des vaisseaux propres et du tannins, dans les musacées. *Ann. Sci. Nat.* 5e sér. **8**: 283–300.
- TRUMBULL, H. L. (1942): Growing rubber in North America. *Ind. Eng. Chem.* **34**: 1328–1335.
- WARNAAR, F. & G. KNOLLEMA (1976): Aromatic and fatty acid components of triterpene esters from Hoya latices. In preparation.