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A COMPARISON OF THE MINERAL RELATIONS OF A HALOPHYTIC HEMIPARASITE AND HOLOPARASITE

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The mineral relations of the halophytic root hemiparasite Odontites verna ssp. serotina and stem holoparasite Cuscuta salina var. major were compared. The xylem hemiparasite Odontites occurs on the upper parts of Dutch Wadden salt marshes attached to the roots of salt excluding monocotyledonous plant species. Cuscuta was sampled in San Francisco Bay (U.S.A.) salt marshes parasitizing Salicornia pacifica of the lower marsh zones. Despite salt accumulation in the host plant Salicornia, the phloem feeder Cuscuta maintains a relatively low Na concentration and a high potassium/calcium (125) ratio. In contrast, the xylem-xylem contact between the strongly transpiring hemiparasite Odontites and its host leads to a 5 to 7-fold increase of the sodium as well as calcium and magnesium content in the hemiparasite shoot tissue. The K/Ca ratio in the attached hemiparasite (8.2) was much lower than in the halophytic holoparasite Cuscuta. The ecological consequences of the differences between the hemi and holoparasite are discussed.

In both holoparasites and hemiparasites contact organs on the shoot and the stem enable the transfer of nutrients from the host to the parasite. In this paper a comparison is made between the mineral relations of a hemiparasite and a holoparasite occurring in salt marshes.

Odontites verna ssp. serotina is an annual, facultative hemiparasite and member of the Scrophulariaceae occurring in salt marshes and beach plains (Ro-ZEMA et al. 1985a). In July 1984, studies were made of populations of Odontites verna ssp. serotina on the Westerkwelder and Beach Plain salt marsh of the Frisian Island Schiermonnikoog (53°29' N, 6°12' E). Using a Scholander's pressure bomb, the total water potential of the plant $\psi_{\rm T}$ was measured at midday (12.00–15.00) and aerial parts of hemiparasite and host plant were collected for mineral analysis (see ROZEMA et al. 1982). On Schiermonnikoog measurements were made on upper marsh sites where individuals of Odontites were present amidst one dominant host species (table 1) making up at least 60% of the total plant cover. By observation of haustoria on root systems Odontites was found

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Odontites verna ssp. serotina in th age values of four replications.	e Wester	kwelder sa	ult-marsh	of Schierr	nonnikoog	g, July 1984. A	Aver-
Species	Minera	l element			ψт	Na host	<u></u>
	mmol/į	gDW	µmol/g	DW		Na paras.	Ca
	K	Na	Ca	Mg			
Host Festuca rubra ssp. litoralis	.540	.140	19.1	26.7	2.20	0.211	
Hemiparasite Odontites verna	.416	.665	50.5	105.3	-2.75		8.2
Host Festuca rubra ssp. litoralis non parasitized	.443	.090	17.9	31.4	-2.04		
Host Juncus gerardii	.580	.175	10.3	23.0	-2.10	0.136	
Hemiparasite Odontites verna	1.138	1.1287	137.2	325.8	-2.84		8.3

Table 1. Mineral composition (mmol or $\mu mol/g$ DW) of the shoot tissue and total water potential of the plant ψ_T (MPa) of two host plant species parasitized and non-parasitized by the hemiparasite *Odontites verna* ssp. *serotina* in the Westerkwelder salt-marsh of Schiermonnikoog, July 1984. Average values of four replications.

to have xylem-xylem contact with at least the following host plant species *Festu*ca rubra spp. litoralis. Juncus gerardii, Agrostis stolonifera and occasionally with Elymus pycnanthus and Trifolium repens.

.178

.085

14.1

11.3

32.8

19.9

-2.17

-.20

.325

.214

Host Juncus gerardii

non parasitized L.S.D. $\alpha = 0.05$

In July 1983 the holoparasite Cuscuta salina var. major (MUNZ & KECK 1959) in a salt marsh of the San Francisco Bay near Palo Alto, (37° 45'N, 122°30'W) San Francisco, was found massively attached to the stem of Salicornia pacifica and to a minor extent to other halophytic species (table 2). Similar midday pressure bomb readings were done as above. Unlike Salicornia the very weak tissue of Cuscuta allowed no proper pressure bomb measurements. The root hemiparasite Odontites absorbs xylem sap with dissolved nutrients from its host by maintaining a significantly more negative water potential than that of its host (table 1, cf KLAREN 1975). This low water potential of the hemiparasite and the continuous flow of xylem water and dissolved nutrients is also related to a high transpiration rate of Odontites even during the dark period to some extent (ROZEMA et al. 1985b). The above monocotyledonous host plants for Odontites were found to be salt-excluding (ROZEMA et al. 1981) halophytes with relatively low Nalevels in the xylem sap. After growth in sand culture with 50 mM NaC1 in the soil moisture the sodium content (mM) of the xylem sap in the hosts, obtained by the pressure bomb technique (ROZEMA et al. 1985b) was 10.0 (Festuca rubra ssp. litoralis), 6.1 (Juncus gerardii) and 2.5 (Elymus pycnanthus) respectively. The potassium content of the xylem sap of all three host species was about 3.5 mM. Nevertheless, the Na-concentration found in the shoot tissue of the hemiparasite exceeds that of the host species by a factor of 5 (Festuca) -7.5 (Juncus). For the ions Ca and Mg a similar high accumulation factor exists, which is in agreement with findings of KLAREN (1975). Potassium concentrations in the hemiparasite were found even lower than that of the host plant species and never more than a factor of 1.5 higher than that of the host plant.

Collected in the lower parts of a San Francisco Bay salt marsh the sodium content of the Salicornia pacifica shoot tissue is much higher (2.96-4.90 mmol Na/g DW) than for the shoot tissue of Festuca rubra ssp. litoralis and Juncus gerardii, growing in Schiermonnikoog salt marshes (0.14-0.175 mmol Na/g DW, table 2). The Na concentration in the parasite Cuscuta tissue was 4-10 times less than that of the host plant tissue. The sodium levels in the stem holoparasite Cuscuta salina may even be lower than that of the root hemiparasite Odontites verna ssp. serotina (tables 1 and 2). There is no accumulation of K, Ca and Mg in Cuscuta relative to the host plant tissue (table 2) either. The content of Mg in Cuscuta tissue is lower than in the host plant, while the potassium content in Cuscuta varies significantly less than in the host. Of the micronutrients, only the Fe content of the holoparasite exceeded that of the host plant species and this is in agreement with data presented by WALLACE et al. 1978. Apparently, the manner of uptake of phloem fluid and dissolved substances by the holoparasite Cuscuta enables the maintenance of low Na concentrations. WALLACE et al. (1978) observed similarly low Na concentrations in the stem holoparasite Cuscuta nevadensis Johnston, parasitizing desert halophytes. This may confirm that the Salicornia phloem and not the xylem is the source of nutrients unlike the relationship between Odontites and its hosts.

According to WOLSWINKEL (1977) phloem feeding parasites have a relatively low Ca and a relatively high K content and consequently a relatively high K/Ca ratio. In accordance with this hypothesis the K/Ca ratio calculated for *Cuscuta* (38.8–218.8) always greatly exceeds the K/Ca ratio determined for *Odontites* (8.2). The data in *table 2* agree with figures presented by Ernst for the root holoparasite *Cistanche lutea* feeding on the halophyte *Halimione portulacoides* (BAU-MEISTER & ERNST 1978). The shoot tissue concentration of Na,K Ca and Mg in *Cistanche* is lower than in the shoot of ist host.

Similar to the shoot hemiparasite Phoradendron villosum, a mistletoe on Quercus lobata (HOLLINGER 1983) and in Rhinanthus (KLAREN 1975), transpiration in Odontites verna ssp. serotina is high and exceeds that of the host plants (ROZE-MA non published). This transpiration rate may contribute to the accumulation of ions in the hemiparasite. Alternatively, in the phloem feeding Cuscuta species, transpiration is low (TER BORG et al 1981). Hemiparasites of the Scrophulariaceae and holoparasites of the Cuscutaceae may be relatively host aspecific (TER BORG et al. 1981). In the particular case of salt marsh environment with different kinds of stress (ROZEMA et al. 1985a), xylem feeding halophytic hemiparasites like Odontites will have a more limited distribution along a gradient of soil salinity than the phloem feeding Cuscuta. Because of the negative waterpotential in the xylem of the host, Odontites must always develop and maintain a more negative water potential by solute uptake or synthesis accompanied by a high transpiration rate (cf table 1, KLAREN 1975). The accumulation of any inorganic ions including the potentially toxic Na⁺ and Cl⁻ is a consequence that limits the distribution of Odontites of the salt marsh to the upper parts in salt-excluding

position (mmol or μ mol/g DW shoot) of the shoot tissue and total water potential of the plant ψ_T (MPa) of four host plant species parasitized by the holoparasite <i>Cuscuta salina</i> var. <i>major</i> in the San Francisco Bay Salt marsh, July 1983. Average values of four iculated at $\alpha = 0.05$ level.
Table 2. Mineral composition (m parasitized and non-parasitized l replications L.S.D. calculated at a

$\begin{array}{ c c c c c c c c c c c c c c c c c c c$		Minera	Mineral element									
K Na Cl Ca Mg Fe Mn P ca 441 2.96 3.90 6.81 27.6 .152 .256 34.10 tsalina .38 .47 .49 3.11 4.6 .792 .232 n.d. 1salina .380 1.31 1.68 7.74 5.6 1.130 .129 49.88 1salina .300 1.31 1.68 7.74 5.6 1.130 .129 49.88 tsalina .239 1.06 1.29 4.89 15.8 .196 .125 44.81 tsalina .299 .119 .72 3.23 .67 37.22 tsalina .56 .13 .69 .21 .203 .087 37.22 tsalina .56 .13 .26 .21 .23 .037 37.22 tsalina .56 .109 .57 .093 .231 .2918 tsalina		mmol/1	g DW		µmol/g	DW	:	,		ψT	Na host	мç
ca .441 2.96 3.90 6.81 27.6 .152 256 34.10 <i>isalina</i> .38 .47 .49 3.11 4.6 .792 .232 n.d. <i>isalina</i> .300 1.31 1.68 7.74 5.6 1.130 .129 49.88 <i>isalina</i> .300 1.31 1.68 7.74 5.6 1.130 .129 49.88 <i>isalina</i> .300 1.31 1.68 7.74 5.6 1.130 .129 49.88 <i>isalina</i> .209 .119 .72 3.23 4.3 .363 .087 37.22 <i>isalina</i> .299 .119 .72 3.23 4.3 .363 .087 37.22 <i>isalina</i> .256 .13 .169 1.17 2.6 .210 .083 40.57 <i>isalina</i> .556 .88 .117 2.6 .210 .083 40.57 <i>isalina</i> .56 .57 .099 .2591 <i>isalina</i> .65 .57 .084 .051 16.52		K	Na	сı	Ca	Mg	Fe	Mn	Р		Na paras.	5
131 1.30 3.4.1 .343 .264 18.06 1salina .300 1.31 1.68 7.74 5.6 1.130 .129 49.88 ifflora .300 1.31 1.68 7.74 5.6 1.130 .129 49.88 ifflora .239 1.06 1.29 4.89 15.8 .196 .125 44.81 1salina .239 .119 .72 3.23 4.3 .363 .087 37.22 1salina .256 .13 .169 1.17 .26 .210 .083 40.57 1salina .256 .13 .169 1.17 .26 .210 .083 40.57 1salina .586 .88 .81 1.19 5.9 .287 .099 25.91 .102 .63 .65 .62 .27 .084 .051 16.52	Salicornia pacifica Parasite Cuscuta salina	.441 .388	2.96 .47	3.90 .49	6.81 3.11	27.6 4.6	.152 .792	.256 .232	34.10 n.d.	-4.21	6.3	124.7
liflora 239 1.06 1.29 4.89 15.8 .196 .125 44.81 <i>tsalina</i> 299 .119 .72 3.23 4.3 .363 .087 37.22 .518 1.17 .841 .94 8.0 .241 .123 129.18 .556 .13 .169 1.17 2.6 .210 .083 40.57 .586 .88 .81 1.19 5.9 .287 .099 25.91 .102 .63 .65 .62 2.7 .084 .051 16.52	Atriplex hastata Parasite Cuscuta salina	.300	4.39 1.31	5.61 1.68	1.90 7.74	34.1 5.6	.343 1.130	.129 .129	18.06 49.88	-2.83	3.4	38.8
.518 1.17 .841 .94 8.0 .241 .123 129.18 .256 .13 .169 1.17 2.6 .210 .083 40.57 .586 .88 .81 1.19 5.9 .287 .099 25.91 .102 .63 .65 .62 2.7 .084 .051 16.52	Frankenia grandiflora Parasite Cuscuta salina	.239 .299	1.06 .119	1.29 .72	4.89 3.23	15.8 4.3	.196 .363	.125 .087	44.81 37.22	-2.42	7.1	92.6
.102 .63 .65 .62 2.7 .084 .051 16.52	Kochia Parasite Cuscula salina Kochia non narasitired	.518 .256 .586	1.17 .13 .88	.841 .169 .81	.94 1.17 1.19	8.0 2.6 5.9	.241 .210 .287	.123 .083 .099	129.18 40.57 25.91	-2.83	9.0	218.3
	$L.S.D.\alpha = 0.05$.102	.63	.65	.62	2.7	.084	.051	16.52	28		

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monocots (Festuca rubra ssp. litoralis, Juncus gerardii, Agrostis stolonifera, Elymus pycnanthus). Phloem feeding halophytic holoparasites like Cuscuta salina and Cistanche lutea (BAUMEISTER & ERNST 1978) absorb only relatively small amounts of sodium and therefore may even occur in the lowest zones of salt marshes, on salt accumulating euhalophytes Salicornia pacifica and Halimone portulacoides. From this ecological point of view Cuscuta salina is expected to be more aspecific with regard to host plant species than Odontites verna ssp. serotina.

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