

NOTES ON THE SPECIES DIVERSITY OF EAST AFRICAN ODONATA, WITH A CHECKLIST OF SPECIES

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Preliminary considerations concerning the species diversity of East African dragonflies and the problems of identifying and using such diversity figures are given. For a detailed approach the basic problem is lack of sufficient data in that area. A checklist of species recorded so far for East Africa is given. Looking at pure species number in relation to area, Uganda is definitely more important for dragonfly diversity than its eastern neighbouring countries. If taking endemism and taxonomic singularity into account, the coastal forests of Tanzania and Kenya are very important too.

INTRODUCTION

Biodiversity is a common and frequently used term nowadays, not only in scientist's or conservationist's circles. Although much has been achieved in documenting biodiversity on a broad scale pattern, questions about existing patterns and the understanding of the multiple factors that play a role in the distribution of biodiversity are still a challenge (GASTON, 2000).

Biodiversity is a useful catchword for political argumentation and for fund raising. Most often no further definition is given, whether it is used as species richness (α -diversity), as species diversity with an adjustment for sampling effects and species abundance or as functional diversity with different weighted species, as key-stone species, endemics, etc. If it is used to compare diversity of different areas, difficulties start as to how to weight such parameters as species richness, endemism, distinctness or taxonomic singularity (VANE-WRIGHT et al., 1991).

The present knowledge of East African Odonata is too small to use more than species richness on a very large geographical scale. Functional diversity assumes a profound knowledge of the species ecology and biogeography and is therefore

difficult to apply to dragonflies of tropical Africa, specially for rain forest species. Often mere species lists are either missing or only very local and/or from short term inventories (e.g. CLAUSNITZER & CLAUSNITZER, 1999; MILLER, 1993).

Despite these shortcomings dragonflies prove to be a very good taxon for biodiversity studies, specially if aiming at applied aspects in the long run. Because they are easy to collect and comparatively well studied taxonomically, ecologically and ethologically dragonflies belong to the priority taxa for biodiversity research and as indicator species (DICASTRI et al., 1992; KIM, 1993; STORK & SAMWAYS, 1995). Dragonflies are already widely used and well accepted in Europe for different indication problems (e.g. CORBET, 1993).

The studies of PINHEY (e.g. 1958; 1961; 1962a; 1969; 1970a; 1974; 1980) provide a firm basis for surveying East African dragonflies. Based on Pinhey's works dragonflies are amongst the very few insect groups in tropical Africa for which comprehensive knowledge on systematics, ecology and distribution for the whole region can be acquired in a reasonable frame of time. In the long run, these data can help to generate biodiversity maps and monitoring programs to aid conservation planning and other management-related topics.

African wetlands in general are argued to belong to some of the most productive ecosystems of the world (KABII, 1996). The survey and conservation of this areas needs to have priority status, because of excessive exploitation and other threats, e.g. changes in water quality due to industrial effluent, agricultural pesticides, siltation and the introduction of alien species (e.g. the Nile Perch *Lates niloticus* or the Water Hyacinth *Eichhornia crassipes*).

The area looked at in this account is restricted to Kenya, Tanzania and Uganda only.

DIVERSITY HOT-SPOTS IN EAST AFRICA

Concerning endemism of forest species in general, the lowland and montane forests of western Uganda, the coastal forests of Kenya and the forests of eastern Tanzania are listed as important areas for East Africa (STUART et al. 1990). Some of these coastal forests have hardly been studied (e.g. NJUNGUNA, 1995) but from preliminary work they appear to support high numbers of endemic species from different taxa (Davenport, pers. comm.). The Eastern Arc forests of Tanzania have been classified amongst the 14 most threatened tropical forest hot-spots worldwide (MYERS, 1988; 1990) and are listed amongst the 25 hot-spots on Earth (MYERS et al., 2000). Some of the Arc forests have experienced a comparatively high attention amongst scientists and conservationists (e.g. CAMMAERTS, 1978; HOCHKIRCH, 1996; MAKUNDI, 1995; NEWMARK, 1999), but the most southern ones especially are still "white spots".

In Kenya, the Kakamega Forest is the last guineo-congolian rain forest patch (see Fig. 1) (KOKWARO, 1988) and is listed as "priority biodiversity conservation

area" for the country (WASS, 1995). There is heavy pressure on the forest in terms of logging and fragmentation (e.g. BROOKS et al., 1999). On the national level, the Kakamega Forest is of considerable importance and gains lots of attention (e.g. ROGO et al., 1999). But comparing the Kakamega Forest with forests in Uganda, it is more or less an impoverished form of the latter.

Concerning wetlands in East Africa, Lakes Victoria and Kyoga in Uganda and the swamps of western Tanzania (Fig. 2) are listed as centres of endemism in STUART et al. (1990).

Kenyan wetlands are more of national importance, since there are only a few of them (CRAFTER et al., 1993). Areas in Kenya, which are important for dragonflies, either in terms of pure species richness or in terms of endemism and taxonomic singularity, coincide with most areas listed as important bird areas (IBA) in BENNUN & NJOROGE (1999).

Unlike Uganda and Kenya the wetlands of Tanzania seem to be hardly studied at all, although almost 10 % of the country's surface area has been estimated to be covered by wetlands (KAMUKALA & CRAFTER, 1993). "Tanzania has extensive but poorly known wetland areas . . . As well as permanent swamps, Tanzania also has seasonally inundated flood plains . . . These (lakes) are all important for their large number of endemic fish and invertebrate species (the details of many of which are poorly known)" (STUART et al., 1990, p. 205).

Uganda is considered as a centre of high biodiversity in Africa (POMEROY, 1993). This is mainly triggered by Uganda's high habitat diversity, ranging from snow-capped mountains to endless swamp and lake areas and its position between the guineo-congolian and the somali-massai savanna region. Most of Uganda's species richness (Tab. 1) is caused by the different forest types and extensive swamp areas (see also HOWARD, 1995b). In Uganda the forest and wetland areas are

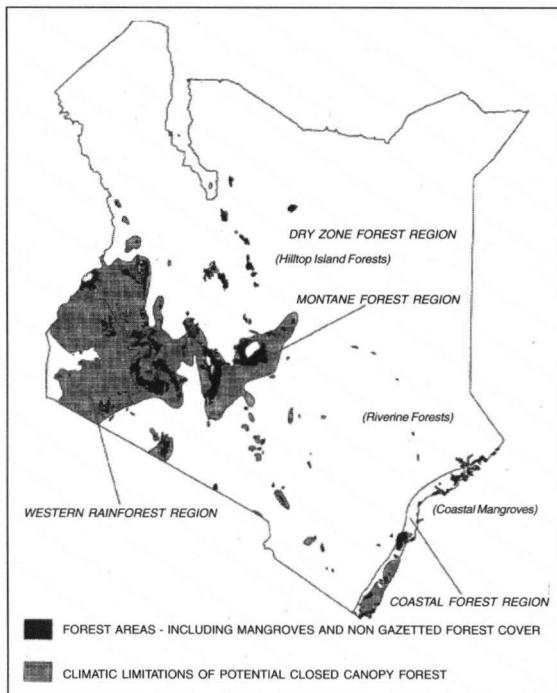


Fig. 1. Distribution of forest and potential forested regions in Kenya (after WASS, 1995).

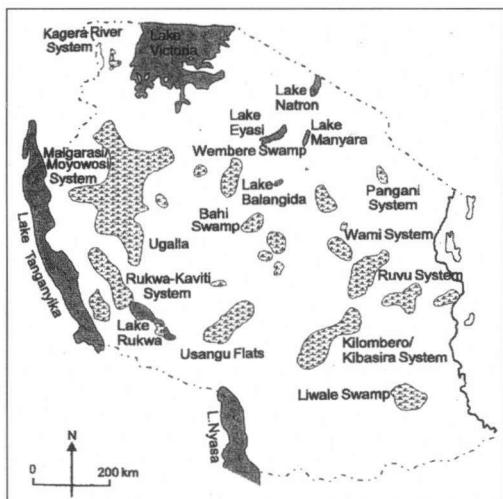


Fig. 2. Map of Tanzania, showing the distribution of swamps and lakes (after KAMUKALA & CRAFTER, 1993).

made for *Pseudagrion bicoerulans*, which can be found up to over 3000 m a.s.l. on most mountain ranges. It is the only species which can be found in the heather zone and in the afroalpine zone, but it is most common along rivers in the montane forest. There are only a few endemics, e.g. *Platycypha amboniensis* from Mt. Kenya, or *Amanipodagrion gilliesi* from the Usambara Mts. Similar observations of the impoverishing dragonfly diversity with increasing altitude have been made on other mountainous areas in Africa as well (e.g. SAMWAYS, 1989; VICK, 1999).

PROBLEMS WITH DIVERSITY

As already mentioned in the introduction, it is not easy to decide, which type of diversity measures one chooses, especially when comparing different areas or countries. In conservation circles, diversity generally means species diversity, which might be corrected for area; better would be an inclusion of endemism level. In most cases the question of what to use depends on the data available and background information concerning ecology and biogeography on the species level.

In a study about the diversity of rheophilous Odonata in southern Spain, FERRERAS-ROMERO (1999) used three components of biodiversity: richness, taxonomic singularity and geographic rarity (endemism). The information on biogeography was taken from the whole geographic range.

In the Sango Bay, Uganda a biodiversity assessment was attempted, integrating field data of different taxa and remote sensing (FULLER et al., 1998). Dragonflies were amongst the taxonomic groups surveyed. Diversity was calculated in terms of

currently surveyed for different taxa and the data are incorporated in the National Biodiversity Data Bank (e.g. FULLER et al., 1998; POMEROY, 1995; REYNOLDS et al., 1999).

Although mountainous areas in Africa are generally considered to be of high conservation value, they are very poor in dragonfly diversity and endemism. A pond on Mt. Elgon (Saito Dam, 2750 m a.s.l.) was inhabited by five species only (*Aeshna e. ellioti*, *Enallagma pseudolongatum*, *E. subfurcatum*, *E. subtile*, *Orthetrum caffrum*) all of them being very common generally. The highest records are

species numbers and species uniqueness using the surveyed area as reference. With this method airstrip ponds gained the highest overall total biodiversity values for dragonflies. Without having detailed checklists of the survey, I assume confidently that most of the airstrip species are not the ones with the highest conservation value, but more or less mainly typical widespread and eurytopic species of savanna ponds. The major habitats in the Sango Bay area are swamp forests, with a very high conservation value (ACREMAN & HOLLIS, 1996; HOWARD, 1995a) and some very interesting dragonfly species (see also MILLER, 1995). When finally ranking the surveyed areas in terms of biodiversity values and producing biodiversity maps, FULLER et al. (1998) referred to the airstrip ponds as "special sites" and focused on the swamp forest and other forest habitats.

These examples show clearly the importance of a broad knowledge of the taxa within the biogeographical region, such as that used by FERRERAS-ROMERO (1999). If the approach in the Sango Bay area had been made for dragonflies only, and without any background knowledge, airstrip ponds would have become the most valuable habitat in terms of biodiversity.

SOME FIGURES FOR KENYA, TANZANIA AND UGANDA

Uganda, even though by far the smallest of the three countries, always comes first in terms of species numbers (Tab. I, Fig. 3). Because of its position between two important biogeographic regions and its extensive wetland and forest areas, it scores for the highest species number. The numbers given in Figure 3 are a bit less than the total number recorded for Uganda (Tab. I), because of species occurring in Uganda and other non-neighbouring countries, e.g. Cameroon. One might argue, that it is a simple bias in study intensity, but this definitely does not apply to Kenya. It is true to some extent for Tanzania, e.g. there are some species with records for Malawi or Zambia and Kenya, but not for Tanzania (see Checklist).

Tanzania, although comparatively poor in species richness (at the present state of knowledge) is the only country with species mentioned in the "Red List of

Table I

Odonates recorded in Kenya, Tanzania and Uganda; neighbouring countries are: Somalia, Ethiopia, Sudan, DRC, Rwanda, Burundi, Zambia, Malawi, Mozambique – [mainly based on literature surveys]

| | km ² | Species | Species common to all 3 countries | Species occurring in at least one of the 3 countries | Species occurring in at least 3 countries or in a neighbouring one |
|----------|-----------------|---------|---|---|---|
| Kenya | 582,646 | 194 | | | |
| Tanzania | 942,444 | 171 | 128 | 296 | 439 |
| Uganda | 236,038 | 249 | | | |

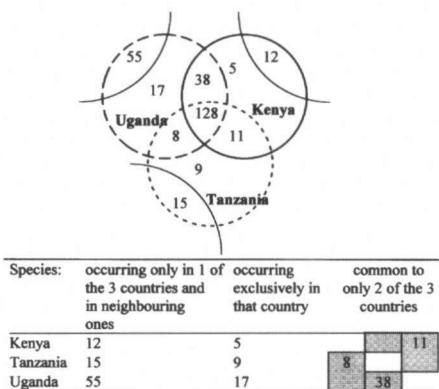


Fig. 3. Odonates recorded only in one or two of the countries Kenya, Tanzania and Uganda, neighbouring countries are: Somalia, Ethiopia, Sudan, DRC, Rwanda, Burundi, Zambia, Malawi, Mozambique [mainly based on literature surveys].

Conservation Action Plan". Of course it is again partially a result of data deficiencies, but taking into account the fact that Tanzania is definitely the worst surveyed country, Uganda is comparatively poor in species with special conservation status.

VICK (1999) lists 180 species for the South-West province of Cameroon and expects a total of at least 200 species. Kenya is about 24 times the area of the province and has a record of at least 194 species (CLAUSNITZER, 1999). Although an increase in Kenya's species number can be confidently expected, it will never reach anywhere near to the area species relation observed in Cameroon. Some areas in Uganda, especially if different habitats along altitudinal gradients are covered, might reach comparative species densities to those observed in Cameroon.

In South Africa 29 dragonfly species (18.7%) are endemic to the area (SAMWAYS, 1999). Looking at the present information available for Kenya, Uganda and Tanzania (Tab. I and Checklist) the degree of endemism is slightly above 5% in each country. But even after an intense survey in all areas, I doubt whether these countries will have a level of endemism comparative to the Cape Region. The percentage of endemics in each country separately from the total dragonfly fauna is even more likely to decrease.

CHECKLIST OF DRAGONFLIES RECORDED IN KENYA, TANZANIA AND UGANDA

Neighbouring countries are only considered, if species have been recorded there and not in Kenya, Uganda and/or Tanzania, but are likely to occur there. Genera

"Threatened Animals" (*Amanipodagrion gilliesi*, *Aeshna meruensis*) and in the "Priority Species: Species of Monotypic Genera confined to one country" list (*Amanipodagrion gilliesi*, *Nepogomphoides stuhlmanni*) (MOORE, 1997). One species (*Coryphagrion grandis*) is listed for Kenya and Tanzania under the "Priority species: taxonomically isolated species" (MOORE, 1997), the given distribution "Uganda" is an incorrect locality interpretation (see PINHEY, 1961) and "Mozambique" is later doubted by PINHEY (1981). No dragonfly species occurring in Uganda is mentioned in the "Status Survey and

and species are listed alphabetically. The survey is based on BARLOW, 1996, CONSIGLIO 1978a, 1978b, DUMONT 1978, FRASER 1953, 1955, GAMBLE 1979, PINHEY 1961, 1962a, 1962b, 1964, 1966, 1967a, 1967b, 1969, 1970a, 1970b, 1971, 1974, 1978, 1981b, 1982, 1984a, 1984b, 1985 and own records. E. Afr: recorded for Uganda, Kenya and Tanzania (not necessarily exclusive); – DRC: Democratic Republic of Congo (former Zaire); – Moz: Mozambique; – Zam: Zambia; – Mal: Malawi; – Eth: Ethiopia; – Som: Somalia; – Sud: Sudan; – Burundi and Rwanda are not listed separately.

| <i>Chlorolestidae</i> | | | |
|--|------------------------------|--|-------------------------|
| <i>Chlorolestes elegans</i> Pinhey, Mal | | | |
| 1950 | | | |
| <i>Lestidae</i> | | | |
| <i>Lestes amicus</i> Martin, 1910 | s.Tanzania, Moz, s.DRC | | |
| <i>L. dissimilans</i> Fraser, 1955 | E. Afr. | | |
| <i>L. ictericus</i> Gerstäcker, 1869 | E. Afr. | | |
| <i>L. ochraceus</i> Selys, 1862 | Kenya, Mal, Zam, | | |
| <i>L. pallidus</i> Rambur, 1842 | E. Afr. | | |
| <i>L. pallidus f. somalicus</i> Förster, 1906 | Kenya, Uganda, Zam | | |
| <i>L. pallidus f. stigmaticus</i> Navas, 1924 | Kenya, Uganda | | |
| <i>L. pallidus f. wahlbergi</i> (Ris, 1921) | Zam | | |
| <i>L. pinheyi</i> Fraser, 1955 | Moz, s.DRC | | |
| <i>L. plagiatus</i> (Burmeister, 1839) | E. Afr. | | |
| <i>L. tridens</i> McLachlan, 1895 | E. Afr. | | |
| <i>L. uncifer</i> Karsch, 1899 | E. Afr. | | |
| <i>L. unicolor aldabrensis</i> Pinhey, 1967 | Tanzania, Kenya (islands) | | |
| <i>L. virgatus</i> (Burmeister, 1839) | E. Afr. | | |
| <i>Protoneuriidae</i> | | | |
| <i>Chlorocnemis abbotti</i> (Calvert, 1892) | Kenya, Tanzania | | |
| <i>C. m. marshalli</i> Ris, 1921 | Mal | | |
| <i>C. marshalli superba</i> Schmidt, 1951 | Uganda, DRC | | |
| <i>C. montana</i> St. Quentin, 1942 | s.Tanzania | | |
| <i>C. nigripes semlikiensis</i> Pinhey 1969 | Uganda | | |
| <i>C. pauli</i> Longfield, 1936 | Uganda, Kenya | | |
| <i>C. wittei</i> Fraser, 1955 | Uganda, s.DRC, Moz, Zam | | |
| <i>Elatoneura glauca</i> (Selys, 1860) | E. Afr. | | |
| | | <i>E. pasquinii</i> Consiglio, 1978 | Eth |
| | | <i>E. tropicalis</i> Pinhey, 1974 | E. Afr. |
| | | <i>Isomecocomnis cellularis</i> Grünberg, 1902 | s.Tanzania |
| | | <i>Prodasineura flavifacies</i> Pinhey, 1981 | n.Zam |
| | | <i>Platycnemididae</i> | |
| | | <i>Allocnemis mitwabae</i> Pinhey, s.DRC 1961 | |
| | | <i>Mesocnemis robusta</i> (Selys, Sud 1886) | |
| | | <i>M. singularis</i> Karsch, 1891 | E. Afr. |
| | | <i>Oreocnemis phoenix</i> Pinhey, Mal 1971 | |
| | | <i>Platycnemis congolensis</i> Martin, 1908 | Uganda, ?Kenya |
| | | <i>P. flavipes</i> Navás, 1924 | Uganda, ?Kenya |
| | | <i>P. nyansana</i> Förster, 1916 | Uganda |
| | | <i>P. xanthopus</i> Navás, 1924 | E. Afr. |
| | | <i>Megapodagrionidae</i> | |
| | | <i>Amanipodagrion gilliesi</i> Pinhey, 1962 | n.Tanzania |
| | | <i>Coryphagrion grandis</i> Morton, 1924 | Kenya, Tanzania |
| | | <i>Coenagrionidae</i> | |
| | | <i>Aciagrion africanum</i> Martin, 1908 | Zam, Moz, s.DRC, Mal |
| | | <i>A. congoense</i> (Sjöstedt, 1917) | Uganda, DRC, Moz |
| | | <i>A. gracile attenuatum</i> Fraser, Mal, Moz, Zam 1928 | |
| | | <i>A. g. gracile</i> (Sjöstedt, 1909) | E. Afr. |
| | | <i>A. hamoni</i> Fraser, 1955 | Uganda, DRC |
| | | <i>A. h. heterostigma</i> Fraser, 1955 | Zam, s.DRC |
| | | <i>A. heterostigma karamoja</i> Pinhey, 1972 | Uganda |
| | | <i>A. s. steeleae</i> Kimmins, 1955 | Zam, DRC |
| | | <i>A. steeleae f. abercornensis</i> Pinhey, 1958 | Zam, ?s.Tanzania |

| | | | |
|--|----------------------------|--|-------------------------|
| <i>A. zambiense</i> Pinhey, 1972 | Zam | <i>P. assegaii</i> Pinhey, 1950 | Zam |
| <i>Agriocnemis aligulae</i> Pinhey, 1974 | Uganda, DRC | <i>P. basicornu</i> Schmidt, 1936 | DCR |
| <i>A. exilis</i> Selys, 1872 | E. Afr. | <i>P. bicoerulans</i> Martin, 1907 | Kenya, n.Tanzania |
| <i>A. forcipata</i> Le Roi, 1915 | Zam, DRC, Sud | <i>P. coelestis</i> Longfield, 1947 | Zam, Mal |
| <i>A. gratiosa</i> Gerstäcker, 1891 | Uganda, | <i>P. coelestis samfyae</i> Pinhey, 1964 | Zam |
| <i>A. inversa</i> Karsch, 1899 | Tanzania, s.Sud | <i>P. commoniae nigerrimum</i> Pinhey, 1950 | E. Afr. |
| | Uganda, Kenya, DRC | <i>P. deningi</i> Pinhey, 1961 | Zam, s.DRC |
| <i>A. palaearia</i> Pinhey, 1959 | Uganda | <i>P. epiphonematicum</i> Karsch, 1891 | DRC, ?Uganda |
| <i>A. pinheyi</i> Balinsky, 1963 | Moz, Zam | <i>P. flavipes</i> Sjöstedt, 1900 | DRC |
| <i>A. pygmaea sania</i> Nielsen, 1959 | n.Kenya, Eth | <i>P. fisheri</i> Pinhey, 1961 | Zam |
| <i>A. victoria</i> Fraser, 1928 | Uganda, Zam | <i>P. gamblesi</i> Pinhey, 1978 | E. Afr. |
| <i>A. zerafica</i> Le Roi, 1915 | Uganda, Kenya, s.Sud, Zam, | <i>P. glaucescens</i> Selys, 1876 | E. Afr. |
| <i>Ceriagrion bakeri</i> Fraser, 1941 | Uganda, Zam, s.DRC, ?Kenya | <i>P. glaucoideum</i> Schmidt, 1936 | DRC |
| <i>C. bidentatum</i> Fraser, 1941 | Uganda, DRC, Mal | <i>P. greeni</i> Pinhey, 1961 | Zam, s.DRC |
| <i>C. corallinum</i> Campion, 1914 | Uganda | <i>P. guichardi</i> Kimmings, 1958 | Kenya, Eth |
| <i>C. glabrum</i> (Burmeister, 1839) | E. Afr. | <i>P. hageni tropicanum</i> Pinhey, 1966 | E. Afr. |
| <i>C. katamborae</i> Pinhey, 1961 | Zam | <i>P. hamoni</i> Fraser, 1955 | E. Afr. |
| <i>C. kordofanicum</i> Ris, 1924 | E. Afr. | <i>P. helena</i> Balinsky, 1964 | Zam, Mal |
| <i>C. moorei</i> Longfield, 1952 | Uganda, Kenya | <i>P. inconspicuum</i> Ris, 1931 | Zam, Mal, s.DRC |
| <i>C. platystigma</i> Fraser, 1941 | Uganda, Kenya, Zam | <i>P. kaffinum</i> Coniglio, 1978 | Eth |
| <i>C. sanguinostigma</i> Fraser, 1955 | Uganda, Zam, s.DRC | <i>P. kersteni</i> (Gerstäcker, 1869) | E. Afr. |
| <i>C. suave</i> Ris, 1921 | E. Afr. | <i>P. kibalense</i> Longfield, 1959 | Uganda, Kenya, Zam, DRC |
| <i>C. whellani</i> Longfield, 1952 | E. Afr. | <i>P. lindicum</i> Grünberg, 1902 | Kenya, Tanzania |
| <i>Enallagma elongatum</i> (Martin, 1907) | E. Afr. | <i>P. makabusiensis</i> Pinhey, 1950 | Zam |
| <i>E. glaucum</i> (Burmeister, 1893) | E. Afr. | <i>P. massaicum</i> Sjöstedt, 1909 | E. Afr. |
| <i>E. longfieldae</i> Fraser, 1947 | Kenya, Uganda | <i>P. melanicterum</i> Selys, 1876 | E. Afr. |
| <i>E. nigridersum</i> Selys, 1876 | E. Afr. | <i>P. niloticum</i> Dumont, 1978 | Eth |
| <i>E. pseudelongatum</i> Longfield, 1936 | E. Afr. | <i>P. nubicum</i> Selys, 1876 | E. Afr. |
| <i>E. sinuatum</i> Ris, 1921 | Tanzania, Zam, DRC | <i>P. quadrioculatum</i> Pinhey, 1964 | DRC |
| <i>E. somalicum</i> Longfield, 1931 | Som | <i>P. r. risi</i> Schmidt, 1936 | DRC |
| <i>E. subfurcatum</i> Selys, 1876 | E. Afr. | <i>P. risi rufocinctum</i> Pinhey, 1955 | Uganda |
| <i>E. subtile</i> Ris, 1921 | E. Afr. | <i>P. rufostigma</i> Longfield, 1947 | Zam |
| <i>E. vansomereni</i> Pinhey, 1955 | n.Uganda | <i>P. salisburyense</i> Ris, 1921 | E. Afr. |
| <i>Ishnura senegalensis</i> (Rambur, 1842) | E. Afr. | <i>P. serrulatum</i> Karsch, 1893 | DRC |
| <i>Mortonagrion stygium</i> (Fraser, 1954) | n.Uganda, DRC | <i>P. sjöstedti beadlei</i> Pinhey, 1961 | Zam, Uganda, ?Kenya |
| <i>Pseudagrion acaciae</i> Förster, 1906 | E. Afr. | <i>P. sjöstedti jacksoni</i> Pinhey, 1961 | Uganda, Zam, s.DRC |
| | | <i>P. sjöstedti pseudosjöstedti</i> Pinhey, 1964 | Tanzania, Moz |
| | | <i>P. s. sjöstedti</i> Förster, 1906 | E. Afr. |
| | | <i>P. sjöstedti wittei</i> Fraser, 1949 | DRC |
| | | <i>P. spernatum gerstaecheri</i> | E. Afr. |

| | | | |
|--|-----------------------|--|--------------------------|
| <i>Karsch</i> , 1899 | | | |
| <i>P. spernatum natalense</i> Ris, | Zam, Mal | <i>C. victoriae</i> (Förster, 1914) | Uganda, DRC |
| 1921 | | <i>C. wittei</i> Fraser, 1955 | n.Zam, s.DRC |
| <i>P. s. spernatum</i> Selys, 1881 | E. Afr. | <i>Platycypha amboniensis</i> (Martin, 1915) | Kenya (central Mts) |
| <i>P. sublacteum</i> (Karsch, 1893) | E. Afr. | <i>P. auripes</i> (Förster, 1906) | Tanzania (coast) |
| <i>P. sublacteum</i> f <i>rusingae</i> Pinhey, 1956 | E. Afr. (L. Victoria) | <i>P. c. caligata</i> (Selys, 1853) | E. Afr. |
| <i>P. s. sudanicum</i> Le Roi, 1915 | Sud, n.DRC | <i>P. l. lacustris</i> (Förster, 1911) | Uganda, w.Kenya, DRC |
| <i>P. sudanicum rubroviride</i> Pinhey, 1955 | Uganda, Zam, DRC, Mal | <i>P. lacustris chingolae</i> Pinhey, 1962 | Kenya, Zam, s.DRC |
| <i>P. sudanicum</i> f <i>vansomerenii</i> Pinhey, 1961 | n.Uganda | <i>P. pinheyi</i> Fraser, 1950 | Tanzania |
| <i>P. symoensii</i> Pinhey, 1967 | Zam | <i>Gomphidae</i> | |
| <i>P. thenartum</i> Fraser, 1955 | DRC | <i>Ceratogomphus pictus</i> Selys, 1854 | s.DRC, Zam |
| <i>P. torridum</i> Selys, 1876 | Kenya, Uganda, DRC | <i>Cinitorgomphus d. dundoensis</i> (Pinhey, 1961) | n.Zam, s.DRC |
| <i>P. torridum</i> f <i>orientis</i> Schmidt, 1951 | n.Kenya | <i>Crenigomphus abyssinicus</i> (Selys, 1878) | Eth |
| <i>P. williamsi</i> Pinhey, 1961 | Zam | <i>C. cornutus</i> Pinhey, 1956 | Zam, ?s.DRC |
| <i>Calopterygidae</i> | | <i>C. denticulatus</i> Selys, 1892 | Eth |
| <i>Phaon iridipennis</i> (Burmeister, 1839) | E. Afr. | <i>C. hartmanni</i> (Förster, 1898) | E. Afr. |
| <i>Umma cincta</i> (Hagen, 1853) | DRC | <i>C. renei</i> Fraser, 1936 | E. Afr. |
| <i>U. declivium</i> Förster, 1906 | Tanzania | <i>Diastatomma rawenzorica</i> Pinhey, 1961 | Uganda |
| <i>U. distincta</i> Longfield, 1933 | Zam, s.DRC | <i>D. selysi</i> Schouteden, 1934 | Zam, DRC |
| <i>U. electa</i> Longfield, 1933 | s.DRC | <i>D. soror</i> Schouteden, 1934 | N.Zam, DRC |
| <i>U. longistigma</i> (Selys, 1869) | DRC | <i>Gomphidia bredoi</i> Schouteden, 1934 | Uganda, Tanzania |
| <i>U. saphirina</i> Förster, 1916 | Uganda, Kenya | <i>G. madi</i> Pinhey, 1961 | Uganda |
| <i>Chlorocyphidae</i> | | <i>G. quarrei confinii</i> Pinhey, 1974 | Moz |
| <i>Chlorocypha aphrodite</i> Le Roi, 1915 | DRC | <i>G. quarrei</i> Schouteden, 1934 | Kenya, s.DRC, Mal |
| <i>C. consueta</i> (Karsch, 1899) | s.Tanzania, DRC | <i>Ictinogomphus ferox</i> (Rambur, 1842) | E. Afr. |
| <i>C. curta</i> (Hagen, 1853) | w.Kenya, Uganda | <i>Lestinogomphus africanus</i> (Fraser, 1926) | Tanzania, Uganda, Kenya |
| <i>C. curta</i> f <i>curta</i> (Hagen, 1853) | n.DRC | <i>L. angustus</i> Martin, 1911 | E. Africa |
| <i>C. cyanifrons</i> (Selys, 1873) | DRC | <i>Microgomphus mozambicensis</i> Moz Pinhey, 1959 | |
| <i>C. frigida</i> Pinhey, 1961 | nw.Zam | <i>M. nyassicus</i> (Grünberg, 1902) | s.Tanzania, n.Zam, s.DRC |
| <i>C. hintzii</i> (Grünberg, 1914) | Uganda | <i>M. schoutedeni</i> (Fraser, 1949) | Uganda |
| <i>C. jacksoni basilewskyi</i> Fraser, 1955 | Burundi | <i>Nepogomphoides stuhlmannii</i> Karsch, 1899 | Tanzania |
| <i>C. j. jacksoni</i> Pinhey, 1952 | w.Uganda | <i>Neurogomphus agilis</i> (Martin, 1908) | DRC |
| <i>C. molindica</i> Fraser, 1948 | Uganda, DRC | <i>N. featheri</i> Pinhey, 1967 | w.Kenya |
| <i>C. molindica hasta</i> Pinhey, 1960 | Tanzania | <i>N. fuscifrons</i> Karsch, 1890 | Uganda |
| <i>C. rubida</i> (Hagen, 1853) | DRC, ?Uganda | <i>N. martininus</i> (Lacroix, 1921) | DRC |
| <i>C. schmidti</i> Pinhey, 1967 | nw.Tanzania, DRC | | |
| <i>C. selysi</i> Karsch, 1899 | ?DRC | | |
| <i>C. tenuis</i> Longfield, 1936 | w.Kenya, Uganda | | |
| <i>C. trifaria</i> (Karsch, 1899) | Uganda, DRC, s.Sud | | |

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|---|-------------------|---|----------------------------|
| <i>N. pinheyi</i> Cammaerts, 1968 | Kenya, Uganda | <i>P. latifasciae</i> Pinhey, 1961 | Mal |
| <i>N. uelensis</i> Schouteden, 1934 | Tanzania, DRC | <i>P. orientalis</i> Fraser, 1957 | Kenya, Uganda |
| <i>N. wittei</i> Schouteden, 1934 | w.Kenya, DRC, Zam | <i>P. symoensis</i> Liefstink, 1969 | s.DRC, Zam |
| | | A e s h n i d a e | |
| <i>Notogomphus butoloensis</i> Fraser, 1952 | Uganda, w.Kenya | <i>Aeshna e. ellioti</i> Kirby, 1896 | E. Afr. |
| <i>N. cataractae</i> Consiglio, 1978 | Eth | <i>A. ellioti usambarica</i> (Förster, 1906) | ne.Tanzania, Kenya Moz |
| <i>N. cottarelli</i> Consiglio, 1978 | Eth | <i>A. meruensis</i> Sjöstedt, 1909 | Tanzania, ? Kenya |
| <i>N. dendrohyrax</i> (Förster, 1906) | Tanzania | <i>A. moori</i> Pinhey, 1981 | nw.Zam |
| <i>N. dorsalis</i> (Selys, 1857) | Uganda, w.Kenya | <i>A. rileyi</i> (Calver, 1892) | E. Afr. |
| <i>N. flavifrons</i> Fraser, 1952 | Kenya, Uganda | <i>A. subpupillata</i> (McLachlan, 1896) | Moz |
| <i>N. immisericor</i> Campion, 1923 | w.Kenya | <i>A. wittei</i> Fraser, 1955 | Zam, s.DRC |
| <i>N. kilimanjaricus</i> (Sjöstedt, 1909) | Tanzania, Kenya | <i>Anaciaeschna triangulifera</i> McLachlan, 1896 | E. Afr. |
| <i>N. lecythus</i> Campion, 1923 | Eth | <i>Anax bangweulensis</i> Kimmins, 1955 | Zam |
| <i>N. leroyi</i> (Schouteden, 1934) | Uganda | <i>A. chloromelas</i> Ris, 1911 | Uganda, Mal, Zam, Moz, DRC |
| <i>N. lujae</i> (Schouteden, 1934) | Uganda, w.Kenya | <i>A. congoiath</i> Fraser, 1953 | DRC |
| <i>N. meruensis</i> (Sjöstedt, 1909) | n.Tanzania, Kenya | <i>A. ephippiger</i> (Burmeister, 1809) | E. Afr. |
| <i>N. nigripes</i> (Sjöstedt, 1909) | Tanzania | <i>A. imperator mauricianus</i> Rambur, 1842 | E. Afr. |
| <i>N. praetorius</i> (Selys, 1878) | Zam, s.DRC | <i>A. parthenope</i> Selys, 1839 | Som |
| <i>N. rueppeli</i> (Selys, 1858) | Eth, Kenya | <i>A. speratus</i> Hagen, 1867 | E. Afr. |
| <i>N. speciosus</i> (Sjöstedt, 1909) | E. Afr. | <i>A. tristis</i> Hagen, 1867 | E. Afr. |
| <i>N. zernyi</i> (St. Quentin, 1942) | s.Tanzania | <i>Gynacantha africana</i> (P.deBeauvois, 1805) | Uganda |
| <i>Onychogomphus bwambae</i> Pinhey, 1961 | Uganda | <i>G. bullata</i> Karsch, 1891 | Uganda, Kenya, Mal |
| <i>O. kitchingmani</i> Pinhey, 1961 | Zam | <i>G. cylindrata</i> Karsch, 1891 | Uganda, Som |
| <i>O. pilosus</i> (Martin, 1911) | Tanzania | <i>G. manderica</i> Grünberg, 1902 | E. Afr. |
| <i>O. quirkii</i> Pinhey, 1964 | Zam | <i>G. ochraceipes</i> (Pinhey, 1960) | Tanzania |
| <i>O. styx</i> Pinhey, 1961 | Uganda, Tanzania | <i>G. sebastopoloi</i> Pinhey, 1961 | Tanzania, Uganda |
| <i>O. supinus</i> Selys, 1854 | Moz | <i>G. sextans</i> McLachlan, 1896 | Zam, DRC |
| <i>O. supinus nigrotibialis</i> Sjöstedt, 1909 | E. Afr. | <i>G. usambarica</i> Sjöstedt, 1909 | Tanzania, Kenya |
| <i>Paragomphus alluaudi</i> (Martin, 1915) | Kenya, Tanzania | <i>G. vesiculata</i> Karsch, 1891 | E. Afr. |
| <i>P. cataractae</i> Pinhey, 1963 | Zam | <i>G. victoriae</i> (Pinhey, 1961) | Uganda |
| <i>P. cognatus</i> (Rambur, 1842) | E. Afr. | <i>G. villosa</i> Grünberg, 1902 | E. Afr. |
| <i>P. elpidius</i> (Ris, 1921) | E. Afr. | <i>G. zuluensis</i> (Balinsky, 1961) | Mal, Moz |
| <i>P. fritillarius f acuminatus</i> (Selys, 1892) | DRC | <i>Heliaeschna cynthiae</i> Fraser, 1939 | Zam, Uganda |
| <i>P. fritillarius f sabicus</i> Pinhey, 1950 | Moz | <i>H. libyana</i> (Fraser, 1928) | Uganda |
| <i>P. genei</i> (Selys, 1841) | E. Afr. | <i>H. trinervulata</i> Fraser, 1955 | Uganda, Tanzania |
| <i>P. magnus</i> Fraser, 1952 | e.Kenya, Moz | <i>H. ugandica</i> McLachlan, 1896 | Uganda, ? Kenya |
| <i>P. nyassicus</i> Kimmins, 1955 | Mal | <i>H. ukereensis</i> Pinhey, 1961 | Tanzania |
| <i>P. pumilio</i> (Rambur, 1842) | n.Kenya, Eth, Sud | C o r d u l i d a e | |
| <i>P. viridior</i> Pinhey, 1961 | Kenya, Uganda | <i>Hemicordulia asiatica</i> Selys, 1878 | Uganda, Mal |
| <i>P. zambeziensis</i> Pinhey, 1961 | Zam | | |
| <i>Phyllogomphus brunneus</i> Pinhey, 1976 | Zam | | |

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|---|------------------|--|--------------------------|
| <i>Phyllomacromia africana</i> Selys, 1871 | Uganda, ?Mal | <i>B. wilsoni</i> Pinhey, 1952 | n.Uganda, s.Sud, E. Afr. |
| <i>P. aureozona</i> Pinhey, 1966 | DRC | <i>Bradinopyga cornuta</i> Ris, 1911 | |
| <i>P. bifasciata</i> (Martin, 1912) | E. Afr. | <i>B. strachani</i> (Kirby, 1900) | E. Afr. |
| <i>P. bispina</i> Fraser, 1954 | Uganda, Zam, DRC | <i>Chalcostephia flavifrons</i> Kirby, 1889 | E. Afr. |
| <i>P. congolica</i> Fraser, 1955 | DRC | <i>Crocothemis brevistigma</i> Pinhey, 1961 | n.Zam |
| <i>P. flavimitella</i> Pinhey, 1966 | Uganda | <i>C. divisa</i> Karsch, 1898 | E. Afr. |
| <i>P. kimminsi junior</i> Pinhey, 1961 | E. Afr. | <i>C. erythrea</i> (Brullé, 1832) | E. Afr. |
| <i>P. melania</i> Selys, 1871 | Uganda | <i>C. sanguinolenta</i> (Burmeister, 1839) | E. Afr. |
| <i>P. monoceros</i> Förster, 1906 | E. Afr. | <i>C. saxicolor</i> Ris, 1919 | Mal, Moz, Zam |
| <i>P. nyanzana</i> Grünberg, 1911 | E. Afr. | <i>Cyanothemis simpsoni</i> Ris, 1915 | DRC |
| <i>P. overlaeti</i> Schouteden, 1934 | Zam, DRC | <i>Diplacodes deminuta</i> Lieftinck, 1969 | Kenya, Uganda, Mal, Zam |
| <i>P. pallidinervis</i> (Förster, 1906) | Kenya | <i>D. lefebvrii</i> (Rambur, 1842) | E. Afr. |
| <i>P. paula</i> Karsch, 1892 | Som, DRC | <i>D. okavangoensis</i> Pinhey, 1976 | Zam |
| <i>P. picta</i> Selys, 1871 | E. Afr. | <i>Eleuthemis buettikoferi</i> Ris, 1910 | Uganda, Tanzania |
| <i>P. pseudoafricana</i> Pinhey, 1961 | Uganda | <i>E. quadriguttata</i> Pinhey, 1974 | Moz |
| <i>P. seydeli</i> Fraser, 1954 | DRC | <i>Hadrothemis camarensis</i> (Kirby, 1889) | Uganda, Kenya |
| <i>P. subtropicalis</i> Fraser, 1954 | Zam, DRC | <i>H. coacta</i> (Karsch, 1891) | Uganda |
| <i>P. sylvatica</i> Fraser, 1954 | w.Kenya, Uganda | <i>H. defecta pseudodeflecta</i> Pinhey, 1961 | Uganda |
| <i>P. unifasciata</i> Fraser, 1954 | Zam, s.DRC | <i>H. defecta</i> (Karsch, 1891) | DRC |
| Libellulidae | | | |
| <i>Acisoma panorpoides ascalaphoides</i> Rambur, 1842 | E. Afr. | <i>H. infesta</i> (Karsch, 1891) | Uganda |
| <i>A. trifida</i> Kirby, 1889 | E. Afr. | <i>H. scabifrons</i> Ris, 1910 | Kenya, Tanzania |
| <i>Aethiothemis bequaerti</i> Ris, 1919 | Mal, Zam, s.DRC | <i>H. versuta</i> (Karsch, 1891) | Zam, DRC |
| <i>A. carpenteri</i> (Fraser, 1944) | Uganda | <i>Hemistigma albipunctata</i> (Rambur, 1842) | E. Afr. |
| <i>A. diamangae</i> Longfield, 1959 | Zam, Mal, s.DRC | <i>Lokia berenice</i> Fraser, 1953 | DRC |
| <i>A. discrepans</i> Lieftinck, 1969 | Zam, s.DRC, Mal | <i>L. circe</i> (Ris, 1910) | DRC |
| <i>A. mediofasciata</i> Ris, 1931 | Mal | <i>L. corydoni</i> Fraser, 1953 | Uganda, Kenya |
| <i>A. palustris</i> Martin, 1912 | ?Som | <i>L. ellioti</i> Lieftinck, 1969 | Zam |
| <i>Aethriamanta rezia</i> Kirby, 1889 | E. Afr. | <i>L. erythromelas</i> Ris, 1910 | DRC |
| <i>Allorhizucha klingi</i> Karsch, 1890 | Zam, DRC | <i>L. gamblesi</i> Lieftinck, 1969 | Zam |
| <i>A. preussi</i> Karsch, 1891 | Uganda, Zam, DRC | <i>L. incongruens</i> (Karsch, 1893) | ?Uganda, DRC |
| <i>Atoconeura b. biordinata</i> Karsch, 1899 | Tanzania, DRC | <i>Macrodipax cora</i> (Kaup, 1867) | Som |
| <i>A. biordinata kenya</i> Longfield, 1953 | Kenya | <i>Malgassophlebia b. bispina</i> Fraser, 1958 | DRC |
| <i>A. biordinata pseudeudoxia</i> Longfield, 1953 | Uganda | <i>M. bispina longistipes</i> (Pinhey, 1964) | nw.Zam |
| <i>A. eudoxia</i> (Kirby, 1909) | Uganda, w.Kenya | <i>Micromacromia camerunica</i> Karsch, 1890 | Kenya, Uganda |
| <i>Brachythemis lacustris</i> (Kirby, 1889) | E. Afr. | | |
| <i>B. leucosticta</i> (Burmeister, 1839) | E. Afr. | | |

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|---|-------------------------|--|--------------------|
| <i>M. miraculosa</i> (Förster, 1906) | Tanzania | <i>O. stemmale</i> (Burmeister, 1839) | E. Afr. |
| <i>Monardithemis flava</i> Longfield, 1947 | Zam | <i>O. taeniolatum</i> (Schneider, 1845) | Som, Sud, n.Kenya |
| <i>Neodythemis fitzgeraldi</i> Pinhey, 1961 | n.Zam, ?s.Tanzania | <i>O. trinacria</i> (Selys, 1841) | E. Afr. |
| <i>Nesciothemis farinosa</i> (Förster, 1898) | E. Afr. | <i>Oxythemis phoenicosceles</i> Ris, 1910 | Uganda |
| <i>N. fitzgeraldi</i> Pinhey, 1955 | n.Zam | <i>Palpopleura deceptor</i> (Calvert, 1899) | E. Afr. |
| <i>Notiothemis j. jonesi</i> Ris, 1919 | E. Afr. | <i>P. jacunda</i> (Rambur, 1842) | E. Afr. |
| <i>N. jonesi auricolor</i> Fraser, 1944 | Uganda | <i>P. jacunda radiata</i> Pinhey, 1982 | Som |
| <i>N. robertsi</i> Fraser, 1944 | Uganda, Kenya, Zam, DRC | <i>P. lucia</i> (Drury, 1773) | E. Afr. |
| <i>Olpogaster fraseri</i> Pinhey, 1955 | Uganda | <i>P. lucia portia</i> (Drury, 1773) | E. Afr. |
| <i>O. fuelleborni</i> Grünberg, 1902 | E. Afr. | <i>Pantala flavescens</i> (Fabricius, 1798) | E. Afr. |
| <i>O. lugubris</i> (Karsch, 1895) | E. Afr. | <i>Parazyxomma flavicans</i> (Martin, 1908) | Uganda |
| <i>Orthetrum abbotti</i> Calvert, 1892 | E. Afr. | <i>Philonomon luminans</i> (Karsch, 1893) | E. Afr. |
| <i>O. africanum</i> (Selys, 1887) | DRC | <i>Porpax asperipes</i> Karsch, 1896 | Zam, DRC |
| <i>O. angustiventre</i> (Rambur, 1842) | Uganda, Kenya, Zam | <i>P. risi</i> Pinhey, 1958 | Moz, Zam |
| <i>O. austeni</i> (Kirby, 1900) | Uganda, Zam | <i>Rhyothemis mariposa</i> Ris, 1913 | Zam, s.DRC |
| <i>O. brachiale</i> (P.deBeauvois, 1805) | E. Afr. | <i>R. notata fenestrina</i> (Rambur, 1842) | E. Afr. (Cl) |
| <i>O. caffrum</i> (Burmeister, 1839) | E. Afr. | <i>R. semihyalina separata</i> (Selys, 1849) | E. Afr. |
| <i>O. caffrum camerunense</i> Gambles, 1959 | DRC (Ruwenzori) | <i>Sympetrum fonscolombii</i> (Selys), 1840 | E. Afr. |
| <i>O. chrysostigma</i> (Burmeister, 1839) | E. Afr. | <i>S. navasi</i> Lacroix, 1921 | Uganda, Kenya, Mal |
| <i>O. guineense</i> Ris, 1910 | E. Afr. | <i>Tetrathemis bijida</i> Fraser, 1941 | Uganda |
| <i>O. hintzii</i> Schmidt, 1951 | E. Afr. | <i>T. camerunensis</i> (Sjöstedt, 1900) | Uganda |
| <i>O. icteromelans cinctifrons</i> Pinhey, 1970 | E. Afr. | <i>T. corduliformis</i> Longfield, 1936 | Uganda, Kenya |
| <i>O. j. falsum</i> Longfield, 1955 | E. Afr. | <i>T. denticauda</i> Fraser, 1954 | Uganda |
| <i>O. j. julia</i> Kirby, 1900 | Uganda, w.Kenya, Mal | <i>T. polleni</i> (Selys, 1877) | E. Afr. |
| <i>O. kristensi</i> Ris, 1911 | Eth | <i>T. ruwenzoriensis</i> Fraser, 1941 | Uganda |
| <i>O. latihami</i> Pinhey, 1966 | DRC | <i>Thermochoria equivocata</i> Kirby, 1889 | Uganda, Zam, Mal |
| <i>O. machadoi</i> Longfield, 1955 | E. Afr. | <i>T. jeanneli</i> Martin, 1915 | Kenya, Tanzania |
| <i>O. macrostigma</i> Longfield, 1947 | Tanzania, s.DRC | <i>Tholymis tillarga</i> (Fabricius, 1798) | E. Afr. |
| <i>O. microstigma</i> Ris, 1911 | E. Afr. | <i>Tramea basilaris</i> (P.deBeauvois, 1805) | E. Afr. |
| <i>O. monardi</i> Schmidt, 1951 | E. Afr. | <i>T. continentalis</i> (Selys, 1878) | E. Afr. |
| <i>O. ransonneti</i> (Brauer, 1865) | Sud | <i>Trithemis aconita</i> Lieftinck, | E. Afr. |
| <i>O. robustum</i> Banlinsky, 1965 | n.Zam | | |
| <i>O. sabina</i> (Drury, 1770) | Sud | | |
| <i>O. saegeri</i> Pinhey, 1966 | Uganda, Zam, DRC | | |

| 1969 | | 1891) | |
|---|------------------|---|---------------|
| <i>T. aenea</i> Pinhey, 1961 | DRC | <i>T. monardi insuffusa</i> | Zam, Mal, Moz |
| <i>T. africana tropicana</i> Fraser, 1953 | DRC | Pinhey, 1970 | |
| <i>T. annulata</i> (P.deBeauvois, 1805) | E. Afr. | <i>T. nuptialis</i> Karsch, 1894 | Uganda, Zam |
| <i>T. anomala</i> Pinhey, 1955 | n.Zam, ?s.DRC | <i>T. pluvialis</i> Förster, 1906 | E. Afr. |
| <i>T. arteriosa</i> (Burmeister, 1839) | E. Afr. | <i>T. pruinata</i> Karsch, 1899 | E. Afr. |
| <i>T. basitincta</i> Ris, 1912 | Uganda, Kenya | <i>T. stictica</i> (Burmeister, 1839) | E. Afr. |
| <i>T. bifida</i> Pinhey, 1970 | Kenya, Zam | <i>T. wernerii</i> Ris, 1912 | E. Afr. |
| <i>T. bredoi</i> Fraser, 1953 | DRC | <i>Urothemis assignata</i> (Selys, 1872) | E. Afr. |
| <i>T. congolica</i> Pinhey, 1970 | DCR | <i>U. edwardsi</i> (Selys, 1849) | E. Afr. |
| <i>T. dichroa</i> Karsch, 1893 | Uganda, Sud, Zam | <i>U. signata aethiopica</i> | Som |
| <i>T. donaldsoni</i> (Calvert, 1899) | E. Afr. | Nielsen, 1957 | |
| <i>T. dorsalis</i> (Rambur, 1842) | E. Afr. | <i>Zygonyx atritibiae</i> Pinhey, 1964 | Zam |
| <i>T. ellenbecki</i> Förster, 1906 | Eth | <i>Z. eusebia</i> (Ris, 1912) | nw.Zam, DRC |
| <i>T. furva</i> Karsch, 1899 | E. Afr. | <i>Z. fallax</i> (Schouteden, 1934) | Uganda |
| <i>T. grouti atra</i> Pinhey, 1961 | ?Uganda, Zam | <i>Z. flavigosta</i> (Sjöstedt, 1900) | Uganda, DRC |
| <i>T. g. grouti</i> Pinhey, 1961 | DCR | <i>Z. natalensis</i> (Martin, 1900) | E. Afr. |
| <i>T. hecate</i> Ris, 1912 | E. Afr. | <i>Z. regalisberti</i> (Schouteden, 1934) | Uganda |
| <i>T. imitata</i> Pinhey, 1961 | Uganda, w.Kenya | <i>Z. torridus</i> (Kirby, 1889) | E. Afr. |
| <i>T. kalula</i> Kirby, 1900 | DRC | <i>Zyxomma atlanticum</i> Selys, 1889 | Uganda |
| <i>T. kirby ardens</i> (Gerstäcker, | E. Afr. | | |

CALCULATING ODONATE DIVERSITY?

The approach of WASSCHER & BOS (2000) in calculating normalised species densities for standard areas ($250 \times 250 \text{ km}^2$: $62,500 \text{ km}^2$) is not feasible for East Africa with the current state of knowledge. Although there are figures for some areas, a simple extrapolation using the formula given by WASSCHER & BOS (2000), shown in parentheses would generate wrong impressions ($S = s \log(N)/\log(n)$; S : estimate of the number of species expected in an area of "N", N : $62,500 \text{ km}^2$, s : number of species known from an area of "n", n : area investigated).

One could calculate the normalised species densities, using the figures given in Table I. Of course I did this and was pleased to see Uganda with 222 species per standard area and Kenya with 161 species per standard area being considerably upgraded. Knowing about the comparatively poor collecting activities in Tanzania, its poor result with 131 species per standard area is not too bad, as it will be definitely increased following to an increase of surveying. But these figures do not say much about the true distribution of dragonflies and their biodiversity status. These figures mainly contribute to science as data for fund-raising and maybe for making political decisions. Kenya for example, with its large desert and semi-desert areas in the north is strongly biased concerning species richness. The coastal areas and the remaining guineo-congolian forest patch in the west (Kakamega

Forest) account for a large part of the total species richness, whereas the dry north is very species poor, but might still contain some very localised and unknown endemics (see also Fig. 1).

The problem with the application of normalised species diversity figures becomes apparent when looking at the simple number of species recorded in the canton Zürich (example given in WASSCHER & BOS, 2000). Seventy species have been observed in an area of 1729 km². Nobody would calculate now the normalised species diversity, (which would be 104), because the total species number for that area is known (80 species). Without this background knowledge, which is the case in many tropical countries, one would be tempted to calculate the normalised species area for comparison with other countries and with this generate wrong impressions for whole countries or areas.

Most often species numbers are corrected for area to obtain a national species density. For countries like Uganda this proves to give "good" results, as the country is small and covered with dragonfly habitats. Kenya would do badly with this method for reasons given above. It is more suitable to compare uncorrected species numbers together with other patterns, e.g. the nationwide distribution of wetlands, the distribution of potential habitats, the number of endangered or rare species, to assess conservation and environmental issues as well.

PURVIS & HECTOR (2000, p. 218) conclude that "biodiversity cannot be reduced to a single number", although "a single number is often what policy-makers want". The authors describe biodiversity as a multitude of facets, of which species richness is only one. Biodiversity considerations on the global scale should be made very carefully (e.g. GASTON, 2000). MYERS et al. (2000) omit invertebrates in general from their documentation, because they are largely undocumented. For East Africa it will still take some time, until the data base allows a comparative approach of species richness on a regional level.

CONCLUDING REMARKS

The most important habitats in terms of diversity for dragonflies in East Africa are mainly rain forest areas, specially the swamp forests in Uganda and the coastal rain forests of Kenya and Tanzania. Although the most common dragonfly habitats, specially in Kenya, are savanna pools and rivers the above-mentioned rain forest areas contain a lot of unique species and, if arguing from the conservational point of view, such habitats are shrinking very fast and experience heavy pressure from outside. For example the Eastern Arc and Coastal Forests of Tanzania and Kenya are cover an area of 2,000 km² today, which is only 6.7% of their original extent (MYERS et al., 2000).

In terms of pure species richness Uganda definitely has a much higher dragonfly diversity than Kenya and Tanzania. But the coastal rain forest areas of Kenya and Tanzania contain some highly localised species, some being endemics and/or of

unique taxonomy (examples given above). The importance of these coastal rain forests is not only seen for dragonflies, for which the database is not that good at the moment, but it is also reflected by other taxa (e.g. WASS, 1995). Over 50% of all threatened forest woody plants in Kenya for example are found in coastal rain forests and 16% are found in the Taita Hills, which are the northernmost Arc Mountains (WASS, 1995). The Kakamega Forest, which belongs to the guineo-congolian rain forest type mainly found in Uganda, is not of major importance when considering on threatened woody plants. Many species found in this forest are rare in Kenya, but not necessarily in the whole geographic region.

Mountainous areas, specially the afroalpine zones are extremely poor in dragonfly species and can be considered as of minor significance in terms of afrotropical dragonfly diversity. This is in contrast to South Africa, where the mountain ranges are considered as sites of special odonatological significance (SAMWAYS, 1999).

The savanna, which cover large areas in Kenya, Tanzania and north Uganda supports a high number of dragonflies, but mainly widespread and eurytopic species. The desert areas in the north of Kenya are poor in species, but also very little known. More surveys could discover new endemics.

As outlined before the major problem is the lack of knowledge on distribution, ecology and biogeography of afrotropical Odonates. For most areas even mere lists of species are not available and this applies not only for dragonflies (e.g. POMEROY, 1995). With the current level knowledge, it is not possible to give applied approaches on East African dragonflies and their diversity, like can be done for South Africa (e.g. SAMWAYS, 1992, 1999), but it should be aimed at. Specially Tanzania is a white spot in terms of odonates (see also PRENDERGAST, 1999). Large areas of Tanzania have never been extensively investigated, if they have been investigated for dragonflies at all. Some species are recorded for Malawi, Zambia, Kenya and/or Uganda, but not for Tanzania (see Checklist), which is probably not the result of a disjunct distribution, but of collecting gaps. In southern Tanzania especially, there appears to be considerable endemism across botanical and zoological taxa in general (Davenport, pers. comm.) and new dragonflies, at least for the country are likely to be found in inventories. But even Kenya seems to be far away from being surveyed to a satisfying level. During a recent trip I found at least 5 species, which have neither been recorded for the country, nor for East Africa before. These are not yet listed in the Checklist.

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