## INFLUENCE OF FATTY ACIDS ON PETUNIA POLLEN GRAINS

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#### SUMMARY

Fatty acids seem to be strong inhibitors of pollen germination in vitro, especially those with a carbon chain of 10 to 12 atoms. With at least one of these acids, namely 10-hydroxy-2-decenoic acid, this property is of practical importance. It prevents the germination of pollen stored by honeybees in their combs. This particular acid is added by the collecting workerbees from the mandibular glands.

10-OH-2 decenoic acid inhibits the respiration of pollen in culture solution, which could be the result of the blockage of one or more enzymes active in pollen-metabolism. In situ on the stigma, however, no inhibition of the germination occurs, probably on account of esterification with glycerides present in the stigma exudate.

### **1. INTRODUCTION**

Pollen of higher plants is stored in high amount by bees and other insects to serve for feeding. When the pollen is brought into the hive it undergoes some changes which can be followed by the change of germination capacity (STANLEY & LINSKENS 1969). The phytocidic substance, with which the bees moisten the pollen while packing (CASTEEL 1912), seems to be a fatty acid, namely the transform of 10-hydroxy-2-decenoic acid (HO-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>6</sub>-CH = CH-COOH) (LU-KOSCHUS & KEULARTS 1968). The fraction with free fatty acids (obtained with florisil column-chromatography) shows if esterified a decrease, and if saponified an increase of inhibiting activity. Of great importance is the question whether the fatty acid containing secretions produced by the bees during pollen-gathering, when spilled on the stigma, are harmful to the reproductive processes of the flower. Besides this it would be interesting to check the influence of fatty acids other than 10-hydroxy-2-decenoic acid on pollen germination.

## 2. MATERIAL AND METHODS

The two self-incompatible clones of *Petunia hybrida*, W166K (incompatibility alleles  $S_1$ ,  $S_2$ ) and T2U ( $S_3S_3$ ) were cultivated as described before (KONAR & LINSKENS 1966a). Pollen germination-tests were carried out in the way described by LUKOSCHUS & KEULARTS (1968).

All chemicals used were of p.a. grade.

All solutions were adjusted to a level of pH 6 to 7 (with NaOH- or HCl-solution) at which pollen germination is optimal. Respiration of pollen was investigated by the Warburg method. To the main compartment was added 1 ml of pollen suspension (5 mg/ml cultivation solution), which was prepared 20 minutes before starting the experiment. In one of the side-compartments 0.5 ml of a 20 percent KOH solution and in the other 0.5 ml of the test substance solution was present. All tests were carried out at  $25^{\circ}$ C.

## 3. RESULTS

3.1. Influence of fatty acids on pollen germination in vitro

To find out if other fatty acids, besides 10-hydroxy-2-decenoic acid, also have an inhibiting action on the germination of *Petunia*-pollen in vitro, a series of saturated fatty acids were tested on these pollen. The results of the tests are summarized in *table 1*.

From this table, we can conclude that practically all fatty acids have an inhibiting action. The intensity of inhibition increases with the length of the carbon chain up to 10–12 atoms and then decreases again. The strongest effect is linked with 10-OH-2-decenoic acid, besides capric and lauric acid.

# 3.2. Influence of 10-hydroxy-2-decenoic acid on the pollen germination in situ

To check the ability of 10-hydroxy-2-decenoic acid to inhibit the pollen germination on the normal germination site of the flower, the stigma surface, 0.01 ml of a 0.01 molar solution of the sodium salt of this fatty acid was pipetted on to the stigma of 10 *Petunia* flowers and 0.01 ml distilled water on the stigma of another 10 flowers. After about 10 minutes the liquid had disappeared and a pollination with W166K pollen (T2U flowers are cross-compatible) was carried out, to find out if any seed formation would take place. After two weeks all flowers had formed mature seeds, so the pollen germination had not been inhibited. That means that the inhibiting principle, normally added to the pollen pellet by the collecting insects, has lost its activity.

3.3. Influence of 10-hydroxy-2-decenoic acid on the respiration of *Petunia* pollen

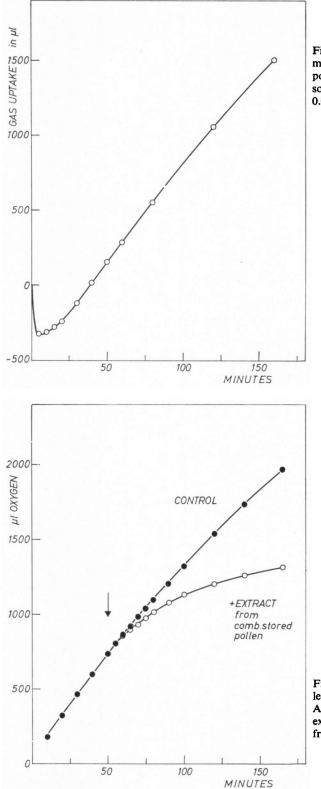
While looking for the mechanism of the action of 10-OH-2-decenoic acid, we studied its influence on pollen respiration. Respiration of pollen of *Petunia* shows in an optimal germination medium a pattern that is characterized by the short depression immediately after wetting and followed by a steady increase of oxygen uptake (*fig. 1*). When pollen respiration is followed after addition of an extract from pollen stored in the hive, one observes depression of oxygen uptake (*fig. 2*). This depression is strong, when extract of the mandibular glands from bees, which produce the inhibition principle (LUKOSCHUS & KEULARTS 1968), is added (*fig. 3*). A  $3.10^{-4}$  molar solution of the sodium salt of 10-hydroxy -2-decenoic acid, added to a pollen suspension that had been respiring for 50 minutes decreased the oxygen uptake almost immediately and even stopped it after about 2 hours. The course of the oxygen-consumption before and after addition of fatty acid solution and water, to test- and control-flask respectively, is shown in *fig. 4*.

## 4. DISCUSSION

It has been known for quite a long time that fatty acids have an influence on the growth of cells. Wyss c.s. (1945) found a fungicidal and bactericidal action

Name of fatty acid         Number         molair concentration $C_{C}$ $C$ $S \times 10^{-6}$ $3 \times 10^{-4}$ $5 \times 10^{-3}$ $2 \times 10^{-3}$ $25 \times 10^{-3}$ $5 \times 10^{-3}$ <t< th=""><th>Table 1. Inhibitory effect of various fatty acids in different concentrations on the germination of <i>Petunia</i> pollen. The figures given are percentages of the germination values of the controls. Because of the low solubilities of some of these fatty acids, their concentrations could not be measured over the entire ranges presented in this table; such instances have been indicated by dashes.</th><th>effect of varic ttion values o tire ranges p</th><th>ous fatty acids of the controls resented in thi</th><th>Inhibitory effect of various fatty acids in different concentrations on the germination of <i>Pet</i> the germination values of the controls. Because of the low solubilities of some of these fatty a over the entire ranges presented in this table; such instances have been indicated by dashes.</th><th>ncentrations on low solubilitie: stances have be</th><th>the germina s of some of sen indicated</th><th>Inhibitory effect of various fatty acids in different concentrations on the germination of <i>Petunia</i> pollen. The figures given are percentages of the germination values of the controls. Because of the low solubilities of some of these fatty acids, their concentrations could not be measured over the entire ranges presented in this table; such instances have been indicated by dashes.</th><th>pollen. The fi their concent</th><th>gures given are rations could ne</th><th>percentages of ot be measured</th></t<>	Table 1. Inhibitory effect of various fatty acids in different concentrations on the germination of <i>Petunia</i> pollen. The figures given are percentages of the germination values of the controls. Because of the low solubilities of some of these fatty acids, their concentrations could not be measured over the entire ranges presented in this table; such instances have been indicated by dashes.	effect of varic ttion values o tire ranges p	ous fatty acids of the controls resented in thi	Inhibitory effect of various fatty acids in different concentrations on the germination of <i>Pet</i> the germination values of the controls. Because of the low solubilities of some of these fatty a over the entire ranges presented in this table; such instances have been indicated by dashes.	ncentrations on low solubilitie: stances have be	the germina s of some of sen indicated	Inhibitory effect of various fatty acids in different concentrations on the germination of <i>Petunia</i> pollen. The figures given are percentages of the germination values of the controls. Because of the low solubilities of some of these fatty acids, their concentrations could not be measured over the entire ranges presented in this table; such instances have been indicated by dashes.	pollen. The fi their concent	gures given are rations could ne	percentages of ot be measured
C- atoms $5 \times 10^{-4}$ $3 \times 10^{-4}$ $5 \times 10^{-4}$ $5 \times 10^{-4}$ $5 \times 10^{-3}$ $2.5 \times 10^{-3}$	Name of fatty	Number				molair co	ncentration			
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	acid	c- atoms	$5 \times 10^{-6}$	$3 \times 10^{-4}$	5 × 10-4	10-*	$1.5 \times 10^{-3}$	$2 \times 10^{-3}$	$2.5 \times 10^{-3}$	$5 \times 10^{-8}$
5       100       100       100       100         6       100       100       100       100       100         8       100       35       0       90       66       66       57         10       35       0       0       0       0       90       66       50       50         11       10       100       100       100       0       0       0       2       55         12       36       0       0       0       0       0       2       50       100       <	butyric acid	4	100	100	100	100	100	100	100	47
6         100         100         100         90           8         100         87         45         0         90           10         35         0         87         45         0         90           11         35         0         0         91         50         100         100         100           14         100         100         100         91         50         100	valeric acid	ŝ	100	100	100	100	100	92	55	16
8 100 87 45 10 35 0 0 0 12 36 0 0 0 14 100 100 91 18 100 100 100 18 100 100 100 100 100	caproic acid	9	100	100	100	8	60		•	
10 35 0 0 12 36 0 0 14 100 100 91 16 100 100 100 18 100 100 100 100 100	caprylic acid	ø	100	87	45	0	0			•
12 36 0 0 14 100 100 91 16 100 100 100 18 100 100 100 100 100	capric acid	10	35	0	0	•				•
14         100         100         91           16         100         100         100           18         100         100         100           10         95         0         0	lauric acid	12	36	0	0	0			•	
16 100 100 18 100 100 10 95 0	myristic acid	14	100	100	91	50	0	0	•	•
id 10 95 0	palmitic acid	16	100	100	100		•		•	•
id 10	stearic acid	18	100	100	100	•	•	•	•	•
10	10-hydroxy-2-									
	decenoic acid	10	95	0	0	0	0	0	0	0

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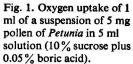
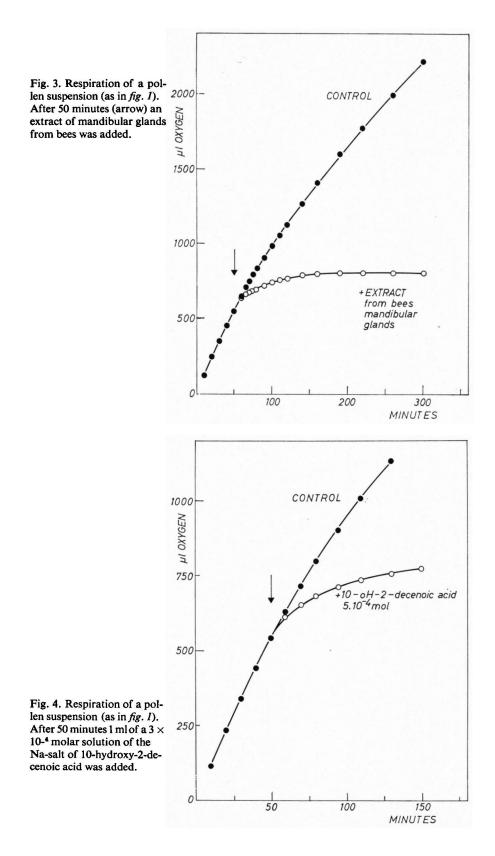


Fig. 2. Respiration of a pollen suspension (as in *fig. 1*). After 50 minutes (arrow) an extract of bee collected pollen from a comb was added.



of fatty acids, especially those with a carbon chain of 11-atoms. ROTHMAN c.s. (1945) established a fungicidal action of hair fat on *Microsporon audouini* and the cause of this action also seemed to be fatty acids with 7 to 11 carbon atoms.

So our results are a fairly good application of the action of fatty acids. Besides the "pollenicidal" action of 10-hydroxy-2-decenoic acid (BUTENANDT & REMBOLD 1957), this substance also inhibits bacterial and fungal growth (BLUM c.s. 1959) and the expansion of artificial leukemic and ascitic tumors (Town-SEND c.s. 1959).

The fact that this acid did not inhibit the pollen tube growth on the stigma and in the style of *Petunia* plants is of great importance. The explanation of this phenomenon could be esterification with glycerides of the stigma exudate of these plants (KONAR & LINSKENS 1966b), for this exudate consists of an oilysubstance free of free fatty acids. So this substance could be a mixture of mono-, di- and triglycerides with many free hydroxylgroups, that could get esterified. If this was also the case with flowers that depend for their reproduction, *e.g.* pollination on insects and especially honeybees, it would be of great importance for these plants as protection against sterilization. There is no contamination of the stigma exudate with pollenicidal substances.

The effect of 10-hydroxy-2-decenoic acid on the respiration does not mean that this substance has a direct influence on this process. It is possible that one or more enzymes, active in pollen metabolism, are blocked and so prevent the course of that special reaction or reactions. Further investigation is in progress.

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